



Comparative Antioxidant and Anti-gout Activities of *Citrullus colocynthis* loaded Fruit Silver nanoparticles with its Ethanolic extract

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Abstract

Background: The biological synthesis of silver nanoparticles (AgNPs) using plant materials is a rapidly developing method with several alternative medical applications. This comparative study of ethanolic fruit extract of *Citrullus colocynthis* (*C. colocynthis*) (EFECC) and synthesized silver nanoparticles (CC-AgNPs) were carried out for antioxidants and anti-gout arthritic activities.

Methods: The AgNPs were synthesized using *C. colocynthis* fruit and its characterization was done by UV-visible spectroscopy, TEM, XRD and FT-IR. The 90% ethanol was used for extract preparation. Antioxidant activity was analyzed by DPPH and the Hydrogen Peroxide (H₂O₂) method. *In vitro* anti-arthritis activity was tested by xanthine oxidase inhibition, protein denaturation and HRBC membrane stabilization assay.

Results: The synthesized CC-AgNPs were confirmed by UV-vis spectroscopy and TEM images displayed spherical shapes with 10-45 nm size range. Furthermore, the functional groups and crystalline structure of CC-AgNPs were determined by FT-IR and XRD analysis. The biosynthesized CC-AgNPs exhibited an excellent free radical scavenging ability than EFECC. In anti-arthritis activity, the CC-AgNPs showed effective inhibition of xanthine oxidase production, protein denaturation, and damaged RBC membranes compared to EFECC.

Conclusion: The antioxidant activities and *in vitro* anti-arthritis assays revealed that CC-AgNPs are better anti-gout agents than EFECC. This research suggested that biosynthesized silver nanoparticles from *C. colocynthis* fruit are an important target in the field of anti-gout drug discovery.

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Introduction

Gout is a dangerous disease affecting people all over the globe and is induced by an accumulation of uric acid in the blood. This causes aggressive rheumatism and extreme pain by triggering the synthesis and deposition of Monosodium Urate crystals (MSU) in the joints ¹. Increased secretion of uric acid and high conception of a purine-rich diet develop hyperuricemia. Gout and hyperuricemia are caused by the oxidation of xanthine and hypoxanthine into uric acid, which is catalyzed by the xanthine oxidase enzyme. High accumulation of urate crystals increased the MSU level in blood due to poor glomerular filtration ². The increased MSU crystals were recently recognized in inflammatory and degenerative tissue reactions involving gouty arthritis through TLR2 and TLR4 receptors ³. The purpose of rheumatoid arthritis treatment is to relieve

symptoms, delay the progression of the disease, and enhance the quality of life for patients. Currently, Disease-Modifying Antirheumatic Medicines (DMARDs) or Non-Steroidal Anti-Inflammatory Medications (NSAIDs) are being replaced by innovative biological agents such as TNF monoclonal antibodies in the treatment of arthritis ⁴. The above therapeutic medications minimize inflammation, joint pains and bone destruction, but their long-term side effects are unclear. Gastric ulcers, hematologic toxicity, hepatic fibrosis and immunological responses are some of the long-term risks of medications ⁵.

Herbal products have been used to cure a lot of diseases since ancient times. Nowadays researchers focused on developing therapeutic drug compounds from plant sources because currently available medicines

either have negative impacts or are extremely expensive. Nature has provided us with a wide variety of herbal plants that are abundantly spread around the world as a source of medicinal substances for the prevention and treatment of a variety of ailments⁶. Many of the important scientific innovations of 21st-century, research and development are believed to be based on nanotechnology. The production of nanoparticles from plant extracts could be used as both reducing and stabilizing agents⁷. The plant extract composition triggers the nanoparticles' functionality. Silver particles were synthesized using multiple approaches, including chemical and physical processes, photochemical reduction, heat evaporation and electrochemical reduction⁸. The dimensions of nanoparticles typically range from 1 to 100 *nanometers* and are an active field of research in nanoscience and nanotechnology⁹. Pharmaceutical industries are currently focusing on developing green mediated nanoparticles with various biomedical applications to reduce toxicity and side effects. Silver nanoparticles (AgNPs) are particularly found in skin ointments and lotions for burns and open wounds¹⁰.

Citrullus colocynthis L. (*C. colocynthis*), which belongs to the Cucurbitaceae family and is generally known as "Colocynth," is found in desert regions of the world¹¹. This plant is native to the Mediterranean region and Asia, however, it can be found in deserts throughout Asia, Europe, and North Africa. The fruits are utilized in Indian traditional medicine to treat a variety of ailments, and people use them to treat joint inflammation, rheumatism, and joint pain¹². According to a literature survey, *C. colocynthis* also has anti-inflammatory, antioxidant, anti-diabetic, antibacterial and anti-cancer properties¹³. The alkaloidal fraction of the fruit extract inhibited the growth of breast and liver cancer cells, providing hope for anticancer potential¹⁴. Therefore, the present comparative study was performed to investigate the antioxidant and anti-gout arthritic effect of synthesized silver nanoparticles from *C. colocynthis* Fruit (CC-AgNPs) and ethanol extract (EFECC).

Materials and Methods

Chemicals used

The chemicals used in this research work such as Ethanol, Bovine Serum Albumin (BSA), Acetyl Salicylic acid and Allopurinol were purchased from SD Fine Chemicals Ltd. DPPH, Hydrogen Peroxide (H₂O₂), Phosphate Buffered Saline (PBS) (pH=7.4), Methanol, and Xanthine oxidase were obtained from Sigma Aldrich.

Collection and extraction of *C. colocynthis* fruit

The *C. colocynthis* fruits were collected from the local region of Madurai, Tamilnadu, India and its identification was confirmed by the Botanical Survey of India (BSI), Coimbatore, India. Collected fruit samples were washed with distilled water to remove impurities and dried in shade. The shade-dried *C. colocynthis*

fruits (500 g) were ground to a fine powder. The fruit extract was prepared using 300 ml of 90% ethanol, added to 100 g of fruit powder and incubated at 37°C for 48 hr in a shaker incubator. The extracts were filtered with three different filters of mesh cloth, filter paper and Whatman no. 1 filter paper and evaporated. This process yielded an ethanolic extract of fruit *C. colocynthis* (EFECC). It was stored in a deep freezer for further analysis.

Synthesis of silver nanoparticles from *C. colocynthis* fruit

1 g of fruit powder was weighed and boiled with 100 ml of distilled water at 60°C for 5 min. After boiling the extract was filtered through Whatman no. 1 filter paper. Then the filtrate was used for silver nanoparticle synthesis. To prepare silver nanoparticles, 10 ml of fruit extract was added into 90 ml of 1 mM silver nitrate solution in a conical flask and was kept in a mechanical shaker. The reduction was started at 1 hr at room temperature, resulting in color changing of aqueous silver nitrate solution indicating the formation of AgNPs. The bio-reduction of silver ions was monitored periodically by measuring UV vis spectra at 250-750 nm¹⁵.

Characterization of nanoparticles

The biosynthesized CC-AgNPs were initially identified visually and then confirmed by double-beam UV-visible spectroscopy (Perkin-Elmer MA, USA). The structural characterization was confirmed by Transmission Electron Microscope (TEM). The crystalline nature of CC-AgNPs was examined using X-ray diffraction (Bruker AXS, Inc., Madison, USA). The bioactive groups in the fruit extract mediated AgNPs were evaluated by Fourier Transform Infrared spectroscopy (FTIR), Perkin-Elmer, MA, USA.

In-vitro antioxidant activity

DPPH radical scavenging assay: In the DPPH free radical scavenging assay, different concentrations (100, 200, 300, 400, and 500 µg/ml) of the test samples (EFECC and CC-AgNPs) were mixed with 1.0 ml of DPPH solution (0.1 mM) in ethanol. Similarly, ascorbic acid was utilized as a known standard, and the results were measured spectrophotometrically at 517 nm after 30 min of incubation¹⁶.

H₂O₂ radical scavenging assay

In this assay, H₂O₂ (40 mM) was prepared in pH=7.4 PBS. Various concentration ranges (100-500 µg/ml) of the test samples (EFECC, CC-AgNPs and ascorbic acid) were separately added to H₂O₂ solution (0.6 ml). The reaction mixture was incubated for 10 min and observed spectrophotometrically at 230 nm¹⁶.

In vitro anti-arthritis activity

Xanthine oxidase inhibitory assay: The xanthine oxidase inhibitory activity of EFECC and CC-AgNPs was assessed by the method of Rifaath¹⁷. In brief, 1 ml of test samples (100-500 µg/ml), 2.9 ml of pH=7.4 PBS and 0.1 ml of xanthine oxidase (0.1 U/ml) were prepared for analysis. The reaction mixture was incubated

at 25°C for 10 min and then a substrate solution of 150 M xanthine was added. The reaction was then inhibited with 1 ml of hydrochloric acid (1 N), and the absorbance was measured by a UV spectrophotometer at 290 nm. As a positive control, allopurinol was used. The xanthine oxidase activity was expressed in percentage inhibition calculated by the following formula:

$$\% \text{ Inhibition} = (1 - A_s/A_c) \times 100$$

Whereas, A_s —Absorbance of test sample, and A_c —absorbance of control.

Protein denaturation study

The effect of EFECC and CC-AgNPs on protein denaturation assay was determined for anti-inflammatory potential and the method was followed by Chaithanya *et al*¹⁸. Different concentrations of EFECC and CC-AgNPs (100-500 µg/ml) were added to 2 ml of 1% BSA and incubated at room temperature for 10 min. Later the mixture was heated in a water bath at 50°C for another 20 min. The inhibition level was measured spectrophotometrically at 660 nm.

HRB membrane stabilization test

In vitro anti-inflammatory activity was determined using the HRBC approach¹⁹. Healthy volunteers' blood samples were drawn and treated with an appropriate amount of sterilized Alsevers solution. The packed cells were separated from the blood solution by centrifugation at 3000 rpm. Isosaline solution was used to wash the packed cells, and isosaline was used to make a 10% v/v cell suspension. The inhibitory activity was examined from this HRBC suspension. Different concentrations of test samples, standard acetylsalicylic acid were mixed individually with 1 ml of PBS, hyposaline (2 ml) and HRBC suspension (0.5 ml). All the above reaction mixtures were incubated at room temperature for 30 min before being centrifuged at 3000 rpm. The hemolysis level of supernatant liquid was determined spectrophotometrically at 560 nm.

Statistical analysis

The obtained results were expressed in Mean±SD. Microsoft Excel 2019 was used to compare the data of the control and test samples. If the p-value is <0.05, the results were considered statistically significant.

Results

Synthesis and characterization of CC-AgNPs

The development of colloidal AgNPs from aqueous extract of *C. colocynthis* fruit when reacted with 1 mM silver nitrate (AgNO_3) solution, slowly changed its color from pale yellow to deep brown color (Figure 1). After 48 hr of incubation, the AgNPs were completely reduced. The synthesized silver nanoparticles were abbreviated as CC-AgNPs. UV-visible spectroscopy is the most useful method for determining the reduction rate of biosynthesized silver nanoparticles. The UV spectra of CC-AgNPs were observed between 250-750 nm and found a strong SPR band at 450 nm (Figure 2), which confirmed the stability of CC-AgNPs. TEM

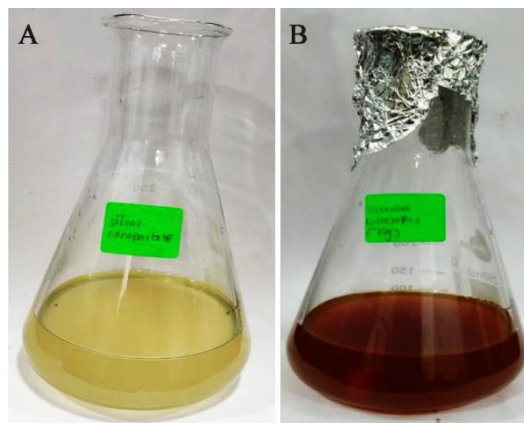


Figure 1. Bio-reduction of Ag ions to AgNPs using *Citrullus colocynthis* fruit extract with 1 mM silver nitrate (A) Initial stage (B) complete reduction after 48 hr.

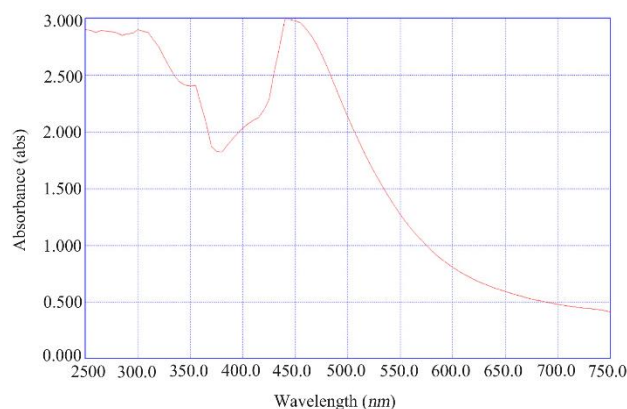


Figure 2. UV-Vis Spectroscopy analysis of CC-AgNPs colloidal solution observed a strong SPR band at 450 nm.

analysis was used to examine the surface morphology of silver nanoparticles derived from *C. colocynthis* fruit extract. The TEM images demonstrated that synthesized CC-AgNPs were mostly spherical and some particles were round in shapes with 10-45 nm size (Figure 3). The crystalline structure of metal nanoparticles was examined using XRD analysis. Figure 4 demonstrated the XRD pattern of *C. colocynthis* fruit synthesized silver nanoparticles. The obtained peaks of CC-AgNPs were displayed at 2θ values of 38.36°, 45.57°, 64.96° and 77.77° assigned to the Bragg's reflection patterns of (111), (200), (220) and (311). According to this analysis, the biosynthesized CC-AgNPs have a Face-Centered Cubic (FCC) structure. The observed peaks were well matched with standard silver structure (JCPDS file Number 84-0713). FTIR analysis was carried out to identify the possible functional groups in the *C. colocynthis* fruit extract-mediated AgNPs. Figure 5 represents the FTIR spectra of CC-AgNPs which have shown peaks at 3333.05 cm^{-1} and 1638.56 cm^{-1} . The strong peak of CC-AgNPs was observed at 3333.05

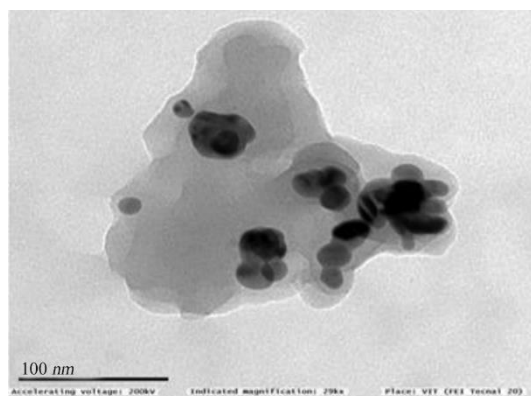


Figure 3. TEM analysis found a surface morphology of synthesized CC-AgNPs as round shaped with 10-45 nm size ranges.

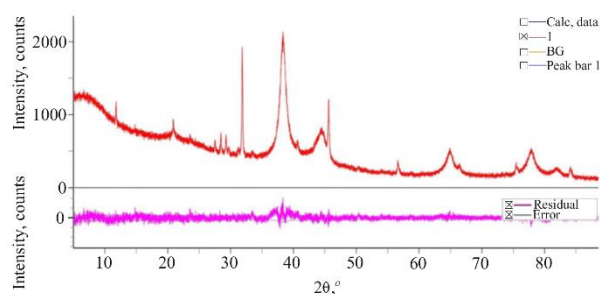


Figure 4. XRD analysis confirmed the crystalline structure of *C. colocynthis* fruit-mediated AgNPs.

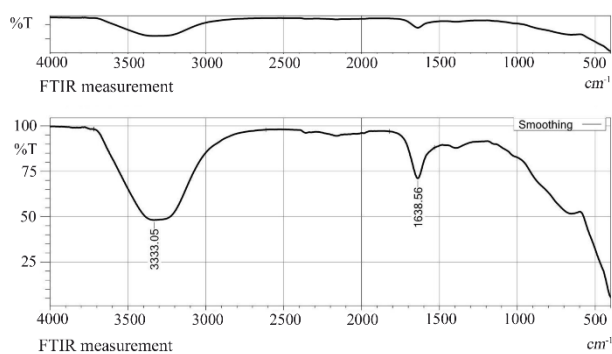


Figure 5. FTIR analysis identified the presence of biochemical components in the fruit extract involved in the CC-AgNPs synthesis mechanism.

cm^{-1} corresponding to -OH stretching of phenolic groups or N-H stretching of primary and secondary amines. The absorption band at 1638.56 cm^{-1} represents C=O stretching of amide or aldehyde or ketone groups. These characterization analyses confirmed the biosynthesized CC-AgNPs.

Antioxidants potential of EFECC and CC-AgNPs

The antioxidant activity was determined by DPPH and H_2O_2 radical scavenging assay by using ascorbic acid as standard. There are five different concentrations

(100, 200, 300, 400 and 500 $\mu\text{g/ml}$) of EFECC, as well as CC-AgNPs which were used to access their free radical scavenging ability. DPPH is a stable synthetic free radical that is easily reduced by fruit extract and silver nanoparticles by absorbing or donating electrons, resulting in the development of the hydrazine molecule, which changes the color of DPPH from purple to yellow. The % inhibition of DPPH was determined by ascorbic acid, EFECC and CC-AgNPs. The free radical scavenging inhibition was gradually increased dose-dependently (Figure 6). This study showed that higher concentrations reported high scavenging ability for EFECC (72.33%), CC-AgNPs (79.9%) and standard ascorbic acid (82.16%). It has been shown that synthesized CC-AgNPs showed significantly higher DPPH radical scavenging ability than EFECC.

The H_2O_2 scavenging activity of EFECC and CC-AgNPs was compared to standard ascorbic acid. This investigation showed increasing scavenging ability with increasing concentration of EFECC and CC-AgNPs (Figure 7). The higher % inhibition was found at 500 $\mu\text{g/ml}$ of EFECC (52.13%) and CC-AgNPs (59.2%). Finally, CC-AgNPs exhibit comparatively better free radical scavenging activity.

In vitro anti-arthritis activity

Xanthine oxidase inhibitory assay: In vitro anti-arthritis activity of EFECC, CC-AgNPs and standard allopurinol was assessed using xanthine oxidase inhibitory activity and the findings were expressed as % inhibition. According to the results of the xanthine oxidase investigation, the percentage inhibition of EFECC and CC-AgNPs showed increase dose-dependently and

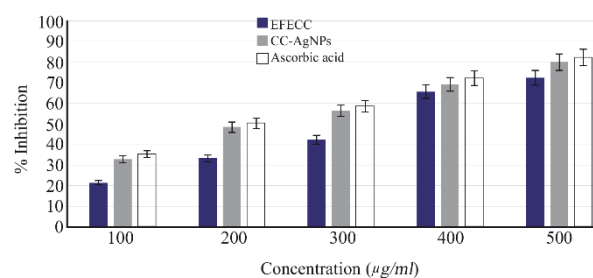


Figure 6. DPPH free radical scavenging potential was observed to be EFECC > CC-AgNPs > ascorbic acid and the %inhibition was increased dose-dependently.

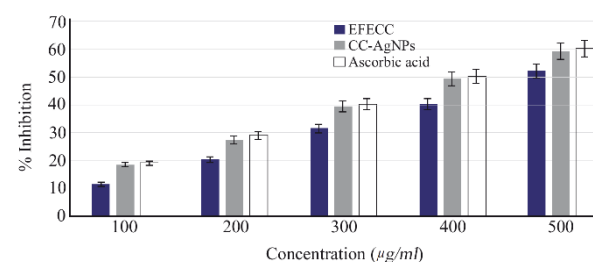


Figure 7. Hydrogen peroxide scavenging activity was observed in EFECC > CC-AgNPs > ascorbic acid and H_2O_2 inhibition also increased dose-dependently.

the higher inhibition was found to be 68.33% and 78.55% respectively at 500 $\mu\text{g/ml}$ concentration (Figure 8). In the present study, CC-AgNPs showed better inhibition activity of xanthine oxidation when compared to EFECC.

Protein denaturation assay

The anti-inflammatory activity of EFECC and CC-AgNPs was measured by the inhibition of protein denaturation assay. In this determination, a better inhibition was observed at a higher concentration (500 $\mu\text{g/ml}$), which also showed concentration-dependent protein denaturation inhibition (Figure 9). Significantly higher inhibition was observed in CC-AgNPs (65.17%) compared to EFECC (53.87%), which were significantly nearer to standard acetylsalicylic acid.

HRBC membrane stabilization assay

The heat and hypotonic saline-induced damage to the RBC membrane was stabilized by EFECC and CC-AgNPs and were comparable with standard acetylsalicylic acid. The EFECC showed a maximum (54.33%) RBC membrane stabilization at 500 $\mu\text{g/ml}$, whereas the CC-AgNPs were found to be 62.3% and standard acetylsalicylic acid showed 63.06% at the maximum concentration (Figure 10). This study also revealed that synthesized CC-AgNPs effectively stabilized the RBC membrane than EFECC.

Discussion

In our present investigation, we described an efficient and ecologically friendly approach for the

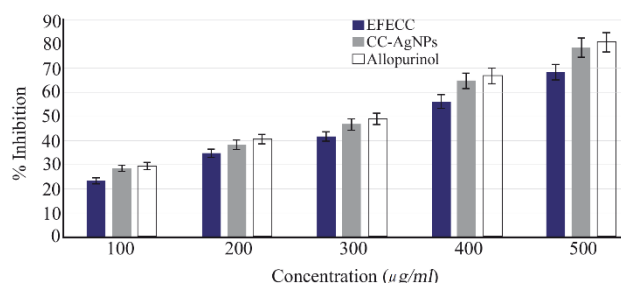


Figure 8. Xanthine oxidase inhibitory analysis proved CC-AgNPs effectively inhibit the xanthine oxidase production similar to Allopurinol standard drug.

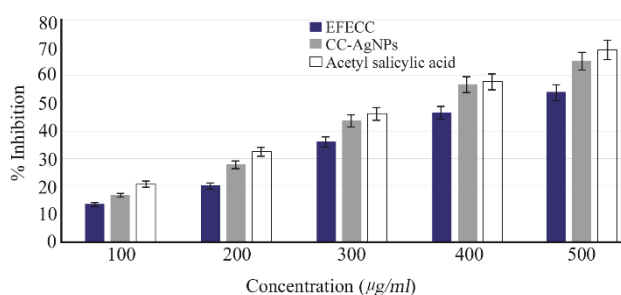


Figure 9. The protein denaturation assay observed the anti-inflammatory activity of Acetyl Salicylic acid >CC-AgNPs> EFECC.

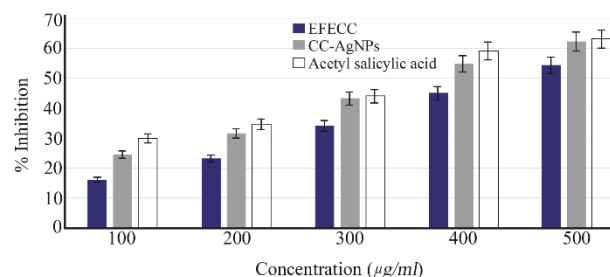


Figure 10. RBC membrane stabilization analysis demonstrated that CC-AgNPs effectively inhibit the damaged RBC membrane when compared to EFECC.

development of silver nanoparticles using the aqueous fruit extract of *C. colocynthis*, which has excellent antioxidant and anti-arthritis properties. UV-visible spectrum detected SPR peak at 450 nm (Figure 2), TEM examination indicated spherical-shaped CC-AgNPs with 10-45 nm size range (Figure 3) and XRD confirmed CC-AgNPs' crystalline structure (Figure 4). FT-IR analysis confirmed the presence of phenolic group, primary and secondary metabolites in the CC-AgNPs (Figure 5). The results of *in vitro* antioxidant activity against DPPH and H_2O_2 radicals after reactions with CC-AgNPs showed better inhibitory action than EFECC (Figures 6 and 7). The improved free radical scavenging activity of green synthesized silver nanoparticles may be attributed to the interaction effect of AgNPs and the bioactive chemicals of plant extract that serve as a capping agent on nanoparticles' surface²⁰. Similarly, AgNPs produced with extracts from *Elephantopus scaber*²¹ and *Coleus aromaticum*²² reported in-cresed free radical scavenging activity.

The CC-AgNPs exhibited excellent anti-gouty arthritic activity through RBC membrane stabilization, inhibition of xanthine oxidase enzymes and protein denaturation. Gouty arthritis therapy aims to achieve a variety of requirements to reduce inflammation, stiffness and should block the excess secretion of uric acid²³. Nanomedicine has considerably improved the pharmacological and pharmacokinetic profiles of traditional drugs used to treat inflammatory arthritis^{24,25}. In the present investigation, the ethanolic fruit extract of *C. colocynthis* and CC-AgNPs effectively inhibited the protein denaturation enzyme and RBC membrane stability. Similarly, the *Phyllanthus tomentosus* leaves extract mediated silver nanoparticles showed strong anti-inflammatory activity through protein denaturation, RBC stabilization and xanthine oxidase analysis²⁶. Previous studies reported that the AgNPs from plant extract possessed excellent antiarthritic activity due to their small size and potential biological system interaction^{27,28}. However, inflammatory cells were able to internalize smaller nanoparticles more easily than their larger particles. The nanoparticles with <400 nm can effectively penetrate into the enlarged cells in rheumatoid arthritis^{29,30}. Compared to previous reports, the

CC-AgNPs with 10-45 nm size ranges potentially inhibited the anti-inflammatory markers.

The current treatment for gout is xanthine oxidase inhibitors, which prevent the secretion of uric acid from purines. Allopurinol is the standard xanthine oxidase inhibitor, that is most commonly recommended to patients for gout treatment³¹. On the other hand, there is a need to create naturally occurring molecules that have the potential to inhibit the xanthine oxidase enzyme to minimize the adverse effects of drugs³². Thus, *C. colocynthis* fruit-mediated AgNPs and its ethanolic extract were investigated against inhibition of xanthine oxidase. This study showed better action of CC-AgNPs with 78.55% inhibitory effect which was significantly similar to standard allopurinol (Figure 8). This clearly demonstrates that the anti-arthritis effect comes from the combination of silver nanoparticles and the stabilizing phytoconstituents in *C. colocynthis* fruit³³. Studies by Mani A *et al*³⁴ have significantly found potent anti-arthritis activity of AgNPs synthesized from *Piper nigrum* extract. The nanoparticles drug delivery system indicated that the nanoparticles were capable of depositing into the affected joints and imposing excellent therapeutic efficacy with acceptable biosafety³³. According to the activity analysis, the CC-AgNPs have the ability to reduce inflammation and treat gout arthritis.

Conclusion

The silver nanoparticles were synthesized from *C. colocynthis* fruit and were confirmed by various spectroscopic techniques. The CC-AgNPs were 10-45 nm in size with spherical shaped, crystalline in nature. The plant source-mediated nanoparticles are non-toxic and ecologically friendly due to the presence of bioactive molecules involved in the reduction process. The therapeutic effect of synthesized CC-AgNPs and ethanol extract of fruit were successfully investigated for anti-oxidant and anti-arthritis activity. The presence of phenols, proteins and amine groups in the fruit extract coated CC-AgNPs exhibited high DPPH and H₂O₂ radical scavenging activity than EFEC. In anti-arthritis assays, CC-AgNPs displayed better inhibition activity of xanthine oxidase enzyme and inflammatory markers than ethanol extract of the fruit. Thus, our findings suggested that biosynthesized CC-AgNPs have anti-gout potential and might play a key role in the development of novel and effective anti-gout therapies.

Acknowledgement

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Conflict of Interest

The authors declare that they have no known competing financial interests in this paper.

References

1. Petchi RR, Vijaya C, Parasuraman S. Anti-arthritis activity of ethanolic extract of *Tridax procumbens* (Linn.) in Sprague Dawley rats. *Pharmacognosy Res* 2013;5(2): 113-7.
2. Alamgeer, Uttra AM, Hasan UH. Anti-arthritis activity of aqueous-methanolic extract and various fractions of *Berberis orthobotrys* Bien ex Aitch. *BMC Complement Altern Med* 2017;17(1):371.
3. Laev SS, Salakhutdinov NF. Anti-arthritis agents: progress and potential. *Bioorg Med Chem* 2015;23(13): 3059-80.
4. Kamel KM, Gad AM, Mansour SM, Safar MM, Fawzy HM. Novel anti-arthritis mechanisms of Polydatin in complete Freund's adjuvant-induced arthritis in rats: involvement of IL-6, STAT-3, IL-17, and NF-κB. *Inflammation* 2018;41(5):1974-86.
5. Arya V, Gupta VK, Kaur R. A review on plants having anti-arthritis potential. *Int J Pharm Sci Rev Res* 2011;7 (2):131-6.
6. Choudhary M, Kumar V, Malhotra H, Singh S. Medicinal plants with potential anti-arthritis activity. *J Intercult Ethnopharmacol* 2015;4(2):147-79.
7. El-Seedi HR, El-Shabasy RM, Khalifa SA, Saeed A, Shah A, Shah R, et al. Metal nanoparticles fabricated by green chemistry using natural extracts: Biosynthesis, mechanisms, and applications. *RSC Adv.* 2019;9(42): 24539-59.
8. Sadeghi B, Gholamhoseinpoor F. A study on the stability and green synthesis of silver nanoparticles using *Ziziphora tenuior* (Zt) extract at room temperature. *Spectrochim Acta A Mol Biomol Spectrosc* 2015;134:310-5.
9. Vijayaraj R, Kumar KN, Mani P, Senthil J, Kumar GD, Jayaseelan T. Green synthesis of silver nanoparticles from ethanolic seed extract of *Acranythes aspera* (Linn.) and its anti-inflammatory activities. *Int J Pharm Ther* 2016;7:42-8.
10. Baraka A, Dickson S, Gobara M, El-Sayyad GS, Zorainy M, Awaad MI, et al. Synthesis of silver nanoparticles using natural pigments extracted from Alfalfa leaves and its use for antimicrobial activity. *Chem Pap* 2017;71(11): 2271-81.
11. Chouaibi M, Rigane K, Ferrari G. Extraction of *Citrullus colocynthis* L. seed oil by supercritical carbon dioxide process using response surface methodology (RSM) and artificial neural network (ANN) approaches. *Industrial Crops and Products* 2020;158:113002.
12. Perveen S, Ashfaq H, Ambreen S, Ashfaq I, Kanwal Z, Tayyeb A. Methanolic extract of *Citrullus colocynthis* suppresses growth and proliferation of breast cancer cells through regulation of cell cycle. *Saudi J Biol Sci* 2021; 28(1):879-86.
13. Kapoor M, Kaur N, Sharma C, Kaur G, Kaur R, Batra K, et al. *Citrullus colocynthis* an important plant in Indian traditional system of medicine. *Pharmacogn Rev* 2020;14 (27):22-7.
14. Hussain AI, Rathore HA, Sattar MZ, Chatha SA, Sarker SD, Gilani AH. *Citrullus colocynthis* (L.) Schrad (bitter

- apple fruit): A review of its phytochemistry, pharmacology, traditional uses and nutritional potential. *J Ethnopharmacol* 2014;8;155(1):54-66.
15. Satyavani K, Gurudeeban S, Ramanathan T, Balasubramanian T. Biomedical potential of silver nanoparticles synthesized from calli cells of *Citrullus colocynthis* (L.) Schrad. *J Nanobiotechnology* 2011;9:43.
16. Sandhiya V, Thirunavukkarasu P, Gomathy B, Sridhar M, Rajeshkumar S, Ravi M, et al. Agnp-Hp synthesized using red marine algae *Halymenia pseudofloresii* and its pharmacological activities. *Annals of the Romanian Society for Cell Biology* 2021;25(6):19433-50.
17. Rifaath M, Kavitha S, Vishnupriya V, Selvaraj J, Gayathri R. An in vitro analysis on the antioxidant and antitumor activity of ethanolic leaf extract of *Mentha Piperita* Linn. *NVEO-NATURAL VOLATILES & ESSENTIAL OILS Journal* NVEO 2021;11:6378-86.
18. Chaithanya MV, Maheswari TU, Rajeshkumar S. Anti-inflammatory and antioxidant activity of lycopene, raspberry, green tea herbal formulation mediated silver nanoparticle. *J Indian Acad Oral Med Radiol* 2021;33(4):397-400.
19. Suganya K, Hari R, Chokkalinagam P, Baskaran P, Maheswaran S, Singh M, et al. Anti-gout arthritic activities of ethanolic and zinc oxide nanoparticle extracts of *Citrullus colocynthis*-An In vitro and Insilico studies. *Annals of the Romanian Society for Cell Biology* 2021;25(3):8020-33.
20. Mohanty AS, Jena BS. Innate catalytic and free radical scavenging activities of silver nanoparticles synthesized using *Dillenia indica* bark extract. *J Colloid Interface Sci* 2017;496:513-21.
21. Kharat SN, Mendhulkar VD. Synthesis, characterization and studies on antioxidant activity of silver nanoparticles using *Elephantopus scaber* leaf extract. *Mater Sci Eng C Mater Biol Appl* 2016;62:719-24.
22. Vilas V, Philip D, Mathew J. Essential oil mediated synthesis of silver nanocrystals for environmental, antimicrobial and antioxidant applications. *Mater Sci Eng C Mater Biol Appl* 2016;61:429-36.
23. Kalpana VN, Payel C, Rajeswari VD. *Lagenaria siceraria* aided green synthesis of ZnO NPs: anti-dandruff, antimicrobial and anti-arthritis activity. *Res J Chemistry Environment* 2017;21(11):14-9.
24. Palani T, Shobha K, Thirunavukkarasu P, Hari R. In vitro and in silico antigout arthritic activities of ethanolic and aqueous stem extracts of *Cissus quadrangularis*-A TLR2 and TLR4 receptor approach. *J Applied Pharmaceutical Science* 2018;8(9):015-22.
25. Viswanathan S, Palaniyandi T, Shanmugam R, Tharani M, Rajendran BK, et al. Biomedical potential of silver nanoparticles capped with active ingredients of *Hypnea valentiae*, red algae species. *Particulate Science and Technology* 2022;40(6):686-96.
26. Salari S, Bahabadi SE, Samzadeh-Kermani A, Yosefzaei F. In-vitro evaluation of antioxidant and antibacterial potential of green synthesized silver nanoparticles using *Prosopis farcta* fruit extract. *Iran J Pharm Res* 2019;18(1):430-5.
27. Govindappa M, Hemashekhar B, Arthikala MK, Rai VR, Ramachandra YL. Characterization, antibacterial, antioxidant, antidiabetic, anti-inflammatory and antityrosinase activity of green synthesized silver nanoparticles using *Calophyllum tomentosum* leaves extract. *Results in Physics* 2018;9:400-8.
28. Ramaswamy M, Solaimuthu C, Duraikannu S. Antiarthritic activity of synthesized silver nanoparticles from aqueous extract of *Moringa concanensis* Nimmo leaves against FCA induced rheumatic arthritis in rats. *Drug Delivery Therapeutics* 2019;9(3):66-75.
29. Hosseinikhah SM, Barani M, Rahdar A, Madry H, Arshad R, Mohammadzadeh V, et al. Nanomaterials for the diagnosis and treatment of inflammatory arthritis. *Int J Mol Sci* 2021;22(6):3092.
30. Zhang S, Wu L, Cao J, Wang K, Ge Y, Ma W, et al. Effect of magnetic nanoparticles size on rheumatoid arthritis targeting and photothermal therapy. *Colloids Surf B Biointerfaces* 2018;170:224-32.
31. Gautam RK, Sharma S, Sharma K, Gupta G. Evaluation of antiarthritic activity of butanol fraction of *Punica granatum* Linn. Rind extract against Freund's complete adjuvant-induced arthritis in rats. *J Environ Pathol Toxicol Oncol* 2018;37(1):53-62.
32. Kiyani MM, Sohail MF, Shahnaz G, Rehman H, Akhtar MF, Nawaz I, et al. Evaluation of turmeric nanoparticles as anti-gout agent: modernization of a traditional drug. *Medicina (Kaunas)* 2019;55(1):10.
33. Biswal B. Standardization protocol development of hydroalcoholic extract of fruits of *Citrullus Colocynthis* against anti-arthritis activity. *International Journal of Green Pharmacy (IJGP)*. 2016 Mar 5;10(1).
34. Mani A, Vasanthi C, Gopal V, Chellathai D. Role of phyto-stabilised silver nanoparticles in suppressing adjuvant induced arthritis in rats. *Int Immunopharmacol* 2016;41:17-23.