

Evaluation of the Bioactive Potential of Secondary Metabolites Produced by a New Marine *Micrococcus* Species Isolated from the Persian Gulf

Hamid Reza Karbalaeei-Heidari ^{1,2*}, Mozhdeh Partovifar ¹, and Mina Memarpour-Yazdi ¹

1. Molecular Biotechnology Lab., Department of Biology, Faculty of Science, Shiraz University, Shiraz, Iran

2. Institute of Biotechnology, Shiraz University, Shiraz, Iran

Abstract

Background: In the present work, a newly isolated marine bacterium, *Micrococcus* sp. MP76, from coastal area of Persian Gulf around Bushehr province, Iran, was identified with the ability to produce bioactive compounds.

Methods: The pigment production was optimized by changing carbon and nitrogen sources in bacterial growth media at 28 °C and 220 rpm for 5 days. Partial purification of the pigment was carried out using suitable solvents.

Results: Maximum pigment extract was achieved (1.4 g/l) when cultured in the medium containing 0.5% (v/v) molasses, 0.5% (w/v) peptone, 1% (w/v) sea salt, 0.01% (w/v) potassium phosphate, and 0.05% (w/v) yeast extract, pH=7.0. Antibacterial effect assessment of the extract against pathogenic bacteria revealed the MIC values in the range of 4.2-7.5 mg/ml depending on different pathogens. The pigment extracted from medium supplemented by molasses and ammonium sulfate had 81% radical scavenging activity, and its IC₅₀ value was 0.28 mg/ml.

Conclusion: The newly isolated strain of *Micrococcus* genus from the Persian Gulf revealed a valuable source to access worth medicinal ingredients when cultured under optimized conditions.

Avicenna J Med Biotech 2020; 12(1): 61-65

Keywords: Antibacterial effect, Antioxidant activity, Bioactive compounds, Marine bacteria, Persian Gulf

* **Corresponding author:**
Hamid Reza Karbalaeei-Heidari,
Ph.D., Molecular Biotechnology
Lab., Department of Biology,
Faculty of Sciences, Shiraz
University, Shiraz, Iran
Tel: +98 71 36137304
Fax: +98 71 32280926
E-mail: karbalaeei@shirazu.ac.ir
Received: 27 Feb 2019
Accepted: 24 Apr 2019

Introduction

Marine bacteria are invaluable resources in providing bioactive compounds ^{1,2}. Marine natural products have significant roles in biomedical research and pharmaceutical industry. In the last decades, a number of bioactive compounds from marine micro-organisms have been identified ³⁻⁶. Although many of these compounds have been used as pharmaceuticals, screening of bioactive agents to find the new chemical structures is still ongoing. The bioactive compounds are typically synthesized during end of the stationary phase of microorganism life cycle. Some of these compounds have been introduced as antibiotics, which play critical roles in surviving and thriving of micro-organisms in bacterial populations or to resist against nutritional stresses ^{7,8}. On the other hand, pigments are one of the most important secondary metabolites which are widely produced by micro-organisms. They are non-toxic, non-carcinogenic and biocompatible and have been welcomed as safe pharmaceuticals. The immune-modulation, antioxidant and anti-carcinogenic properties of carotenoid compounds, as the most important identified pigments, have been reported as well ^{9,10}.

Actinobacteria are a class of gram-positive bacteria which are well known as pioneers in the production of

a wide variety of useful secondary metabolites such as antitumor agents, antibiotics, antioxidants, pigments and enzymes ^{11,12}. Among actinobacterial genera, *Micrococcus* has been reported to contain significant potential to produce a large number of useful compounds. For instance, canthaxanthin (4',4'-diketo-13-carotene) and α - and β -carotene derivatives were purified and characterized from *Micrococcus roseus* (*M. roseus*) ^{13,14}. Moreover, crude pigments from *Micrococcus luteus* (*M. luteus*) revealed antibacterial effects against *Staphylococcus* sp., *Klebsiella* sp., and *Pseudomonas* sp. ¹⁵.

According to this background, the aims of the current study can be summarized as isolation and characterization of a pigment producing bacterium from Persian Gulf, optimization of pigment production using different carbon and nitrogen sources, and finally evaluation of its antibacterial and antioxidant potentials.

Materials and Methods

Isolation of pigment producing bacteria

Pigment producing bacteria were isolated from water samples collected from coastal area of Persian Gulf around Bushehr province, Iran. Micro-organisms were precultured on nutrient agar plates (Difco) containing

3% (w/v) sea salt at 30 °C, pH=8.0 for 5 days. Primary culture medium was composed of 0.3% (w/v) yeast extract, 3% (w/v) sea salt and 0.5% (w/v) casein hydrolysates (pH~8.0). The effect of different carbon and nitrogen sources (0.5%) on pigment production yield was investigated. To produce secondary metabolites, 1% (v/v) seed culture of the strain MP76 was inoculated into production medium and incubated at 28 °C and 220 rpm for 5 days.

Biochemical and molecular characterization of the strain MP76

Morphological and molecular features of the isolate (MP76) were characterized. Biochemical tests were carried out according to the prevalent procedures described by Quinn *et al* ¹⁶. For phylogenetic classification, the 16S rRNA gene sequence of the strain MP76 was amplified with forward primer of HRK1 (5'-AC-TCCTACGGGAGGCAGCAG-3'), and reverse primer of HRK2 (5'-TGACGGGCGGTGTGTACAAG-3') using a thermocycler (Biomerta, USA). After nucleotide sequencing, alignment analysis and phylogenetic tree construction were carried out by MEGA 6.06 software ¹⁷.

Optimization of bacterial growth and pigment production

To investigate the effect of pH and sea salt concentration on the strain MP76 growth, biomass dry weight was measured in production media containing 1-10% w/v sea salt and the pH range of 5.0-9.0. To prepare the extract from the bacterial cells, biomass collected from 5-day culture was suspended in acidic methanol (methanol containing 4% v/v HCl 1N), vortexed gently for 5 min, then separated by centrifugation at 8000 x g for 10 min. The obtained crude pigment extract was dried at 40 °C and used for further analysis.

Study on the pigment extract bioactivities

Antibacterial effect: Antibacterial potential of the pigment extract was investigated by Well Diffusion method (WD), as described by Valgas *et al* ¹⁸, with a slight modification. Antibacterial effects of the pigment extract were studied against the pathogenic bacteria such as *Pseudomonas aeruginosa* (*P. aeruginosa*) (ATCC 27853), *Escherichia coli* (*E. coli*) (ATCC 25922) and *Staphylococcus aureus* (*S. aureus*) (ATCC 25923). The discs were impregnated with 20 µl of the pigment extract (6.3 mg/ml) and placed on the agar plates. After 16-18 hr incubation at 37 °C, diameter of the clear zone of growth inhibition was measured.

Minimum Inhibitory Concentration (MIC) is defined as the lowest concentration of antibacterial compound which prevents visible growth of a bacterium after 24 hr of incubation at 37 °C.

Three pathogenic bacteria were cultivated in NB for 18 hr at 37 °C, then MTT solution (10 µl, 0.5 mg/ml) was added into each well and incubated at 37 °C for 4 hr. After adding 100 µl of the solubilizing buffer and incubation in the dark for 24 hr, the absorbance at 590

and 630 nm was recorded using an ELISA reader. All experiments were carried out in triplicate.

Antioxidant assay

DPPH scavenging activity was determined as described by Memarpour-Yazdi *et al* ¹⁹, with slight modification. Briefly, 660 µl of 0.1 mM DPPH was mixed with 40 µl of the pigment extract in a final concentration of 0.4 mg/ml. After mixing vigorously for 2 min, the mixture was kept at 37 °C for 30 min. Then, the absorbance of the mixture was recorded at 517 nm using a UV-Vis spectrophotometer (Shimadzu, UV. 120-02). DPPH Radical Scavenging Activity (RSA) was determined based on the following equation:

$$\text{RSA (\%)} = [(A_{\text{control}} - A_{\text{sample}}) / A_{\text{control}}] \times 100$$

Trolox is commonly employed as a standard or positive control in antioxidant assays.

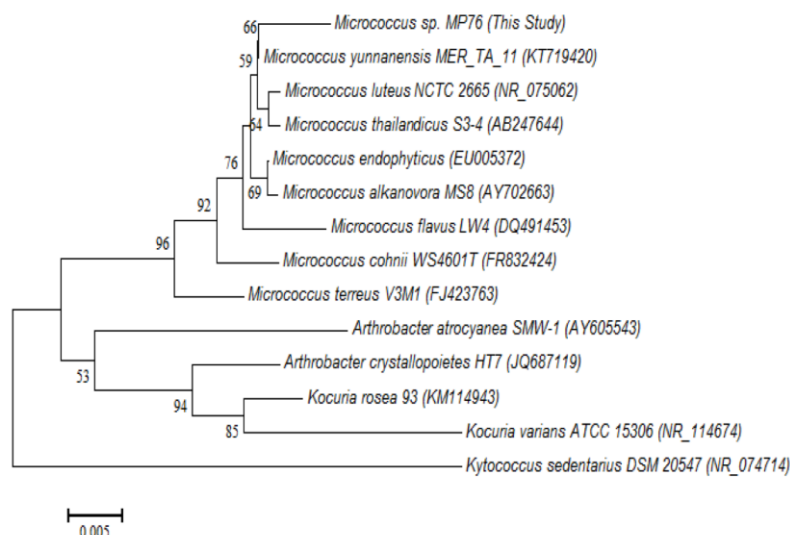
Results and Discussion

Isolation and characterization of the pigment producing bacterium

Screening and production of bioactive compounds and finding the economical culture medium has gained much attraction among researchers. Accordingly, in this study, Persian Gulf coast area was studied to isolate and characterize an efficient bacterium. The strain MP76 was isolated from coastal area of Bushehr province and characterized as a pigment producing bacterium. Isolate MP76 was identified as a gram-positive coccus, with yellow color colonies. Some biochemical characteristics of the isolate (MP76) are determined and summarized in table 1. For the molecular characterization, a consensus sequence of 911 bp fragment of its 16S rDNA gene was identified and deposited in the GenBank database under accession number KT804695. The sequence alignment revealed that the isolate MP76 belongs to *Micrococcus*, a genus of bacteria in the Micrococcaceae family and named as *Micrococcus* sp. strain MP76. The phylogenetic tree (Figure 1) showed that *Micrococcus* sp. strain MP76 is closely related to *Micrococcus yannanensis* (KT719420), a gram-positive bacterium isolated from a plant ²⁰.

Table 1. Biochemical characterization of the isolated strain MP76

Test	Result
Gram staining	Gram positive
Morphology	Circular
Pigment	Yellow
Indole production	-
Lysine dehydroxylase	+
H ₂ S production	-
Urease	-
Oxidase	+
Catalase	+
Coagulase	+
Gelatinase	-
Sucrose/Glucose fermentation	-/-

Figure 1. Phylogenetic tree of *Micrococcus* sp. MP76.

Optimization of bacterial growth and biomass production

The maximum biomass production was observed at the pH range of 6.0-8.0 and the highest was obtained to be 2.7 g/l at pH 7.0 (Figure 2A). Moreover, the noticeable reductions in biomass weight to about 0.95 and 0.8 g/l were observed at pH=5.0 and 9.0, respectively. The highest growth rate was obtained at 0-3% salt (2.8 g/l at 1% sea salt), although it was dropped significantly at higher than 5% (Figure 2B). Effect of different carbon and nitrogen sources revealed that maximum methanolic extract amount was obtained in molasses medium, which was 0.58 g/l, while it declined to 0.18, 0.2, 0.23 and 0.26 g/l in the medium containing olive oil, starch, sucrose and glucose, respectively (Figure 3A). However, the ratio of methanolic extract to biomass, defined as the pigment production yield, was relatively unchanged. As shown in figure 2B, the highest methanolic extract was 1.4 g/l in the medium supplemented by peptone. Although the methanolic extract for peptone and yeast extract were much higher than that of other nitrogen sources, no remarkable difference in pigment production yield was shown among the tested nitrogen sources (Figure 3B).

Pigment characterization

Micrococcus sp. strain MP76 produced a yellow pigment, which was dissolved in methanolic extraction of the biomass. The absorption spectrum of chloroform extract of the pigment had three maximum wavelengths of 418, 437 and 448 nm, showing the most similarity with carotenoids. The maximum wavelengths of the obtained pigment were similar with carotenoid produced by *M. luteus*¹⁵. It has been reported that *M. luteus*, isolated from sea water, was able to produce a yellow pigment with antibacterial activity against *Klebsiella*, *P. aeruginosa* and *S. aureus*¹⁵.

Determination of bioactivities of the extracted pigment

Antibacterial activity: Primary antibacterial tests indicated that the pigment extracts derived only from cul-

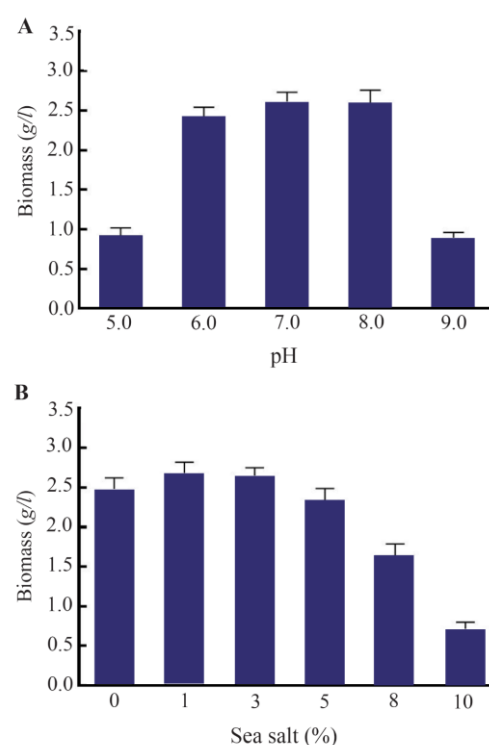


Figure 2. Effect of pH (A), and sea salt concentration (B) on biomass dry weight.

ture containing yeast extract and peptone as nitrogen sources have shown positive results (Figure 4), while among various carbon sources, all pigment extracts had almost the similar inhibitory effects at the same concentrations against *E. coli*, *P. aeruginosa* and *S. aureus* (data not shown). Accordingly, the active pigment extracts were selected to determine IC₅₀ and MIC values. The MIC values of the methanolic pigment extract were obtained to be 4.2, 5.0 and 7.5 mg/ml against *S. aureus*, *P. aeruginosa*, *E. coli*, respectively. Furthermore, the IC₅₀ values against *S. aureus*, *P. aeruginosa*,

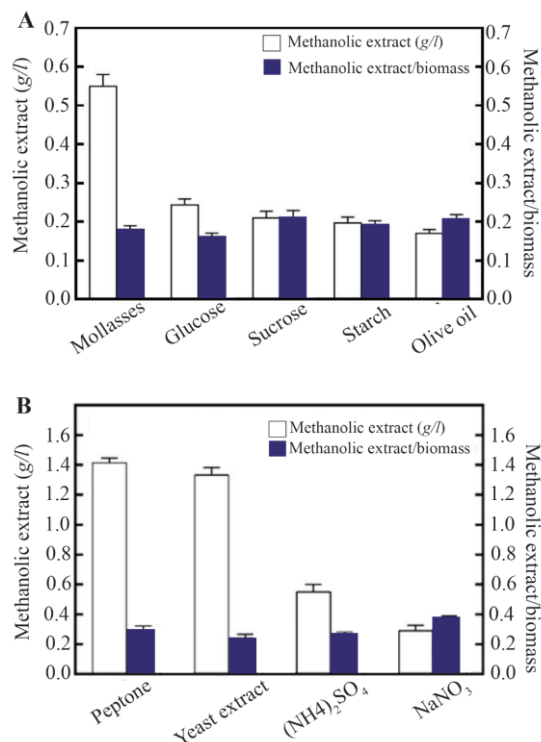


Figure 3. Effect of carbon (A), and nitrogen sources (B) on methanolic extract and the ratio of methanol extract to biomass.

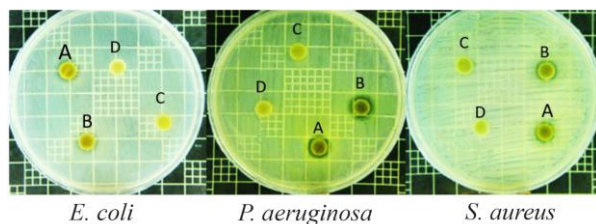


Figure 4. Antibacterial effects of pigment extract obtained from different nitrogen sources including peptone (A), yeast extract (B), (NH₄)₂SO₄ (C), and NaNO₃ (D)

E. coli were 3.4, 4.8, 4.8 mg/ml, respectively. The pigment extract revealed the highest antibacterial activity against the drug resistant hospital bacterium, *S. aureus*. It has been reported that ethyl acetate extract of the supernatant, produced by a species of *Micrococcus* isolated from Bamboo tree waste, showed the MIC value of 256 µg/ml against *S. aureus*²¹. Moreover, Duraikannu *et al* reported almost similar antibacterial effect from the crude extract of a new marine soil isolate, *Streptomyces gancidicus* VITSD1²².

Investigation of the bacterium susceptibility to different antibiotics revealed that *Micrococcus* sp. strain MP76 was susceptible to all tested antibiotics except for streptomycin (Figure S1)²³. Based on the observed resistance to streptomycin, it can be suggested that the antibiotic produced by *Micrococcus* sp. MP76 might belong to aminoglycoside family of antibiotics that includes streptomycin.

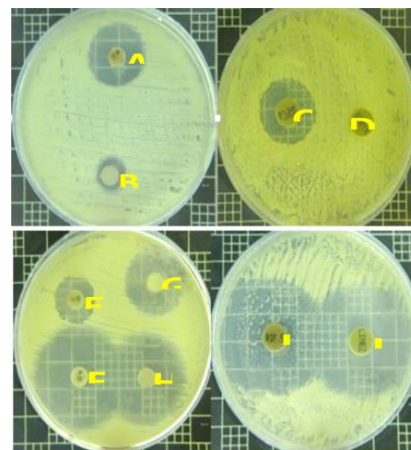


Figure S1. Susceptibility of *Micrococcus* sp. MP76 to different antibiotics including: Ciprofloxacin (A); Chloramphenicol (B); Daptomycin (C); Streptomycin (D); Vancomycin (E); Penicillin (F); Trimethoprim (G); Erythromycin (H); Rifampicin (I) and Linezolid (J).

Table 2. DPPH scavenging capacity (%) of the obtained ethanol extract from different carbon and nitrogen sources

Source	RSA (%) ^a	Trolox equivalent concentration (mM)
Carbon sources		
Molasses	81.68	36.47
Glucose	51.39	22.35
Sucrose	50.29	21.76
Starch	22.00	8.82
Olive oil	19.50	7.64
Nitrogen sources		
Peptone	61.28	27.05
Yeast extract	66.11	28.82
(NH ₄) ₂ SO ₄	81.68	35.88
NaNO ₃	43.35	18.82

a) The average of standard deviations was ± 2%.

Antioxidant activity

Ability of the pigment extracts obtained from different carbon and nitrogen sources for DPPH Radical Scavenging Activity (RSA) was determined at the concentration of 0.4 mg/ml. As illustrated in table 2, among carbon sources, media supplemented by molasses and olive oil showed the highest and the least RSA values, respectively. RSA value of molasses was 1.6-, 3.7- and 4.2-fold higher than that of sucrose, starch and olive oil, respectively. RSA value of ammonium sulfate was 1.8 and 1.2-fold more than that of sodium nitrate and yeast extract, respectively. The pigment extracted from media supplemented by molasses and ammonium sulfate had the highest RSA value (81%) and their IC₅₀ value was 0.28 mg/ml. The most effective antioxidant activity of the pigment extract was obtained in molasses medium, which can be considered as a cost-effective and valuable source to obtain this bioactive compound from the strain MP76. Sugar beet molasses is composed of sucrose (near 48%) and some

other compounds in small scales including nitrogen, sulfur and other minerals such as calcium, potassium, chloride and oxalate ²⁴.

Conclusion

Based on the data of this research, it can be concluded that untapped offshore resources such as the Persian Gulf have the potential to isolate bacteria with the ability to produce bioactive compounds under optimized conditions, and research to identify these bacteria as well as purification of active compounds can be a way to discover the effective antibacterial compounds.

Acknowledgement

The authors would like to thank the research affairs of Shiraz University, Shiraz, Iran. This work was supported in part by a grant for Scientific Research from the Iran National Science Foundation (INSF) under contract number 93035486.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Schinke C, Martins T, Queiroz SC, Melo IS, Reyes FG. Antibacterial compounds from marine bacteria, 2010-2015. *J Nat Prod* 2017;80(4):1215-1228.
- Kiuru P, D'Auria MV, Muller CD, Tammela P, Vuorela H, Yli-Kauhaluoma J. Exploring marine resources for bioactive compounds. *Planta Med* 2014;80(14):1234-1246.
- Trindade-Silva AE, Lim-Fong GE, Sharp KH, Haygood MG. Bryostatins: biological context and biotechnological prospects. *Curr Opin Biotechnol* 2010;21(6):834-842.
- Singh R, Sharma M, Joshi P, Rawat DS. Clinical status of anti-cancer agents derived from marine sources. *Anti-cancer Agents Med Chem* 2008;8(6):603-617.
- Palomo S, González I, de la Cruz M, Martín J, Tormo JR, Anderson M, et al. Sponge-derived kocuria and micrococcus spp. as sources of the new thiazolyl peptide antibiotic kocurin. *Mar Drugs* 2013;11(4):1071-1086.
- Solanki R, Khanna M, Lal R. Bioactive compounds from marine actinomycetes. *Indian J Microbiol* 2008;48(4):410-431.
- Hibbing ME, Fuqua C, Parsek MR, Peterson SB. Bacterial competition: surviving and thriving in the microbial jungle. *Nat Rev Microbiol* 2010;8(1):15-25.
- Chao L, Levin BR. Structured habitats and the evolution of anticompetitor toxins in bacteria. *Proc Natl Acad Sci USA* 1981;78(10):6324-6328.
- Asker D, Ohta Y. Production of canthaxanthin by extremely halophilic bacteria. *J Biosci Bioeng* 1999;88(6):617-621.
- Surekha PY, Dhanya P, Sarath Josh M, Pradeep S, Benjamin S. *Micrococcus luteus* strain BAA2, a novel isolate produces carotenoid pigment. *Electronic J Biol* 2016;12(1):83-89.
- Fiedler HP, Bruntner C, Bull AT, Ward AC, Goodfellow M, Poterat O, et al. Marine actinomycetes as a source of novel secondary metabolites. *Antonie Van Leeuwenhoek* 2005;87(1):37-42.
- Bull AT, Stach JE. Marine actinobacteria: new opportunities for natural product search and discovery. *Trends Microbiol* 2007;15(11):491-499.
- Ungers GE, Cooney J. Isolation and characterization of carotenoid pigments of *Micrococcus roseus*. *J Bacteriol* 1968;96(1):234-241.
- Cooney JJ, Marks HW, Smith AM. Isolation and identification of canthaxanthin from *Micrococcus roseus*. *J Bacteriol* 1966;92(2):342-345.
- Umadevi K, Krishnaveni M. Antibacterial activity of pigment produced from *Micrococcus luteus* KF532949. *Int J Chem Analytical Sci* 2013;4(3):149-152.
- Quinn GP, Keough MJ. Experimental design and data analysis for biologists. United kingdom: Cambridge University Press; 2002.557 p.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 2013;30(12):2725-2729.
- Valgas C, Souza SMD, Smânia EF, Smânia Jr A. Screening methods to determine antibacterial activity of natural products. *Brazil J Microbiol* 2007;38(2):369-380.
- Memarpoor-Yazdi M, Asoodeh A, Chamani J. A novel antioxidant and antimicrobial peptide from hen egg white lysozyme hydrolysates. *J Funct Foods* 2012;4(1):278-286.
- Zhao GZ, Li J, Qin S, Zhang YQ, Zhu WY, Jiang CL, et al. *Micrococcus yunnanensis* sp. nov., a novel actinobacterium isolated from surface-sterilized polyspora axillar roots. *Int J Syst Evol Microbiol* 2009;59(Pt 10):2383-2387.
- Sharma SC, Shovon MS, Jahan MS, Asaduzzaman A, Khatun B, Yeasmin T, et al. Antibiotic sensitivity and antibacterial activity of *Micrococcus* sp SCS1. *Res Rev BioSci* 2012;6:304-310.
- Duraikannu D, D Chandrasekaran S, N Selvakumar J, Vaithilingam M. A preliminary study of in vitro antioxidant and antibacterial activity of *Streptomyces gancidicus* VITSD1 isolated from marine soil. *Curr Bioact Compd* 2014;10(4):292-297.
- Amabile-Cuevas CF, Cárdenas-García M, Ludgar M. Antibiotic resistance. *Ame Sci* 1995;83(4):320-329.
- Curtin LV. Molasses-general considerations. *Molasses in Animal Nutrition* 1983;1-10.