

Development and Comparison of In-house Line Probe Assay (LiPA) and SYBR Green Real-time PCR Regarding the Detection of Periodontal Pathogens

Mohammad Soleimani* and Mohammad Reza Zolfaghari

Department of Microbiology, Qom Branch, Islamic Azad University, Qom, Iran

Abstract

Background: Periodontal disease, which can become a chronic condition, is an inflammatory disease that upsets the soft and hard structures supporting the teeth. The aim of the present study was to design and develop an in-house Line Probe Assay (LiPA), to detect putative periodontitis-related bacterial pathogens, and compare it with SYBR Green Real-time PCR.

Methods: The LiPA method was launched using biotinylated 16s rRNA universal primers and specific probes for each of the five bacteria including *Aggregatibacter actinomycetemcomitans*, *Prevotella intermedia*, *Tannerella forsythia*, *Porphyromonas gingivalis* and *Treponema denticola*. For this, optimized quantities of the primers and specific probes were dotted onto nylon membrane stripes in a defined pattern. Hybridization was performed between the probes and the single-stranded biotinylated PCR products. The stripes were developed *via* biotin-streptavidin reaction. Ultimately, the analytical and diagnostic sensitivity and specificity of the in-house LiPA was evaluated and compared with SYBR Green Real-time PCR.

Results: The detection limit of the LiPA was 2760 copies of targeted genes. In testing analytical specificity, only signals corresponding to the specific biotinylated products were produced. The calculated diagnostic sensitivity of the LiPA for the five bacterial targets ranged from 96.4 to 100%, whereas the diagnostic specificity was between 90.9 and 100%. Comparing the results, no noticeable difference ($p=0.4795$) was observed between the two methods.

Conclusion: To screen periodontal pathogens, a simple, inexpensive and accurate method is desirable. The in-house LiPA, having advantages such as high specificity and sensitivity, and the ability to detect five major periodontal pathogens, offers the option of evaluating samples without the need for a post-PCR platform.

Avicenna J Med Biotech 2019; 11(1): 80-87

Keywords: Periodontitis, Real-time PCR, Sensitivity and specificity

Introduction

As a polymicrobial infection, periodontal disease disturbs tissues supporting and investing the teeth and alveolar bone, and can eventually bring about dental loss. The chronic periodontal diseases are common in the middle-aged and the elderly, while the aggressive periodontal diseases are rampant particularly among youths (30-years-old)¹⁻³. The prevalence of periodontal diseases in America and developing countries has been estimated about 35 and 50-55%, respectively⁴.

During the formation of periodontal disease, the normal oral microbiome shifts gradually from facultative gram-positive bacteria into anaerobic gram-negative ones^{1,5,6}. *Aggregatibacter actinomycetemcomitans* (Aa) (formerly *Actinobacillus actinomycetemcomitans*) and *Porphyromonas gingivalis* (Pg) are the major pathogenic bacteria species in the etiology of periodontal diseases, whereas the normal oral flora related to the

disease are *Prevotella intermedia* (Pi), *Tannerella forsythia* (Tf) and *Treponema denticola* (Td)⁷⁻⁹.

It is necessary to evaluate microbial composition in human periodontal pockets for the diagnosis and treatment of periodontal diseases¹⁰. Currently, there are various methods to detect the odontopathogen bacteria in saliva or plaque¹¹. In a former study, conventional PCR and Real-time PCR were developed and compared for detection of Aa and Tf. The SYBR green Real-time PCR was tenfold more sensitive than the conventional PCR⁷. Multiplex PCR and Real-time PCR are powerful and efficient tools in detecting concurrently bacterial strains in periodontal samples^{12,13}. However, multiplex PCR has several drawbacks, including complexity, low amplification efficiency, and poor universality¹⁴. Likewise, the use of Real-time PCR was also complicated with expensive equipment

* Corresponding author:
Mohammad Soleimani, Ph.D.,
Department of Microbiology,
Qom Branch, Islamic Azad
University, Qom, Iran
Tel: +98 251 7780001
Fax: +98 2517780001
E-mail:
soleimanidor@yahoo.com
Received: 26 Aug 2017
Accepted: 24 Dec 2017

and reagents¹⁵. As a diagnostic molecular method, Line Probe Assay (LiPA) is based on a DNA strip that allows for a rapid and simultaneous identification of the target DNA sequences. In-house and commercially available line probe assays are currently being used to detect the members of the *Mycobacterium tuberculosis* Complex (MTC) and their drug-resistant strains¹⁶⁻¹⁹, and also for genotyping Noroviruses²⁰, human papillomavirus (HPV) in cervical scrapes²¹ and Hepatitis C virus²²⁻²⁷. The aim of the present study was to design and apply the Line Probe Assay for a rapid screening of several prominent oral pathogens including Aa, Pi, Tf, Pg and Td bacteria.

Materials and Methods

Strains and DNA extraction

The genome of wild-type bacteria of Aa (ATCC 700 685D-5), Tf (ATCC 43037D-5), Td (ATCC 35405D-5), Pg (ATCC 33277D-5) and Pi (ATCC 25611D-5) was used as positive control. The bacterial species including *Staphylococcus aureus* (ATCC 25923), *Bacillus subtilis* (ATCC 6051), *Shigella sonnei* (ATCC 92 90), *Escherichia coli* (*E. coli*) (ATCC 25922), *Enterococcus faecalis* (ATCC 29212), *Pseudomonas aeruginosa* (AT CC 27853), *Klebsiella pneumoniae* (ATCC 7881), *Yersinia enterocolitica* (PTCC 1480), *Yersinia pseudotuberculosis* (PTCC 1244), *Proteus vulgaris* (PT CC 10 79) and *Citrobacter freundii* (PTCC 1499) were used as negative control. The bacterial genomes were extracted using EZ-10 spin Column Genomic DNA Kit (DNPTM kit (Bio Basic Inc., Canada)); the purity of the genomes was subsequently evaluated. The extracted genomes were quantified using a Picodrop Spectrophotometer (Picodrop, UK) according to the manufacturer's instructions.

Primers and probes

NCBI deposited sequences of 16S rRNA gene of the target bacteria were downloaded and aligned by CLC Main Workbench software version 7.5 (accession numbers are not shown). A pair of broad range universal bacterial primers was selected to amplify a 466-bp region: Forward primer 5'-TCCTACGGGAGGCAGCA GT-3' and reverse primer 5'-GGACTACCGGGTAT CTAATCCTGTT-3'²⁸.

The 466 bp region of the 16S rRNA alignment was designated to design specific probes for each of the five target bacteria through the use of CLC Main Workbench software version 7.5 (Table 1). Confirming the specificity of the designed probes, the sequences were compared against the NCBI nucleotide databases.

The biotinylated and non-biotinylated forms of the universal primers and the probes containing 5' poly (T) sequences were commercially synthesized.

PCR and cloning

PCR experiments were conducted in a volume of 25 μ l, consisting of 1X PCR buffer (Fermentas, GmbH, Germany), 2 mM MgCl₂, 0.2 mM dNTPs, 0.5 μ M of the non-biotinylated universal bacterial primers, 50 ng of genomic DNA (each of the five target bacteria) and 1 unit Taq DNA polymerase (Fermentas, GmbH, Germany). Amplification was carried out for 35 cycles in the PCR program as follows: Initial denaturation at 94°C for 4 min, denaturation at 94°C for 30 s, annealing temperature at 54°C for 30 s, extension time at 72°C for 1 min and final extension for 10 min. The 466 bp PCR products of 16S rRNA genes (each of the five bacteria) were cloned in plasmid vector by use of InsTA clone PCR cloning kit (Fermentas, Lithuania) according to the manufacturer's instruction. The recombinant plasmids were transformed into CaCl₂ competent *E. coli* Top10 F' strain. The confirmed recombinant plasmids containing 16S rDNA insert of Aa, Tf, Td, Pg and Pi were named pTZ-agg, pTZ-tan, pTZ-tre, pTZ-por and pTZ-pre, respectively. The plasmids were subjected to PCR same as above except for the fact that biotinylated primers were used. The biotinylated PCR products were stored at -20°C until use in LiPA experiment.

Preparation of the LiPA strips

The probes (Table 1), the biotinylated universal primers (Conjugate control) and the PCR products from non-biotinylated universal primers (Amplification control), were prepared with concentrations of 2.5 pmol/ μ l; they were then spotted onto SensiBlot™ Plus Nylon membranes (Fermentas GmbH, Germany) employing a 96-well Dot-Slot Blotter (Cleaver, UK) according to the manufacturer protocol. Fixation of the spotted membranes was done through baking at 80 °C for 30 min. Furthermore, the spots were UV cross-linked with 100-150 μ J/cm² on the nylon membranes. The prepared membranes were sliced into 60×7 mm strips and stored at -20°C until use.

Prehybridization, hybridizations, and color development

The prepared stripes were prehybridized in 2X SSC buffer as blocking solution [1X SSC: 0.15 M NaCl (Hi-Media, India) and 0.015 M Sodium Citrate (Appli-Chem GmbH, Germany)] and 0.1% SDS (Cinnagen, Iran) at 50 °C for 30 min. Each prehybridized strip was transferred to a 14 ml polystyrene conical bottom test

Table 1. The specific probes for each of the five target bacteria used in the study

Bacterium	Probe name	Position	Probe sequence (5'-3')	Length (bp)	Reference
<i>A. actinomycetemcomitans</i>	Agg-p	617-632	CTTTAGGGAGGGGTAG	16	This study
<i>P. gingivalis</i>	Por-p	736-751	TTGCCATACTGCGACT	16	This study
<i>P. intermedia</i>	Pre-p	662-667	GTGCACGCAACGTATG	16	This study
<i>T. forsythia</i>	Tan-p	655-670	TATAGATGAAGTAGGC	16	This study
<i>T. denticola</i>	Tre-p	674-689	ATCACGGAGGGGAAAC	16	This study

Table 2. Different mixtures of the positive control prepared in order to determine the specificity

	pTZ-agg	pTZ-por	pTZ-pre	pTZ-tan	pTZ-tre
Mixture 1	10 ng/ μ l	10 ng/ μ l	0	0	0
Mixture 2	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l	0	0
Mixture 3	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l	0
Mixture 4	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l	10 ng/ μ l

tube containing 2 ml hybridization buffer (2X SSC 0.1% SDS). Adding 10 μ l of the single-stranded biotinylated PCR products, hybridization was accomplished in an orbital shaker at 50°C and 80 rpm for 1-2 hr. Washing the strips twice with 2X SSC and 0.1% SDS for 5 min at 22°C was followed by washing twice with 1X SSC and 0.1% SDS for 15 min at 42°C. The hybridized strips were developed via Biotin Chromogenic Detection Kit (Fermentas GmbH, Germany) according to the manufacturer instructions. All steps were performed at 22°C with moderate shaking on an orbital platform shaker. Primarily, the strips were treated with 2 ml blocking/washing buffer for 5 min and then blocked in 2 ml of the blocking solution for 30 min. Next, the strips were immersed in 2 ml of diluted Streptavidin-AP conjugate for 30 min. After washing with blocking/washing buffer and incubating with detection buffer for 10 min, enzymatic reaction and color development was performed. The strips were incubated in 2 ml of fresh substrate solution [5-Bromo, 4-Chloro, 3-Indolylphosphate (BCIP) and Nitro-blue-tetrazolium (NBT)] in a dark place. The blue-purple precipitate was produced after 20-30 min of incubation. After discarding the substrate solution, the strips were rinsed with Milli-Q water to stop the enzymatic reaction. Lastly, the strips were interpreted through inspecting the blue-purple bands corresponding to controls and bacterial species.

Analytical sensitivity and specificity

A 10 ng/ μ l concentration (2.76×10^9 copies) of the positive control plasmids (pTZ-agg, pTZ-tan, pTZ-tre, pTZ-por and pTZ-pre) was used to prepare a 10-fold serial dilution of each. The dilutions were subjected to PCR amplification using the biotinylated primers. The specific PCR products were detected through the use of the designed in-house LiPA as previously mentioned. The last dilutions demonstrating relevant blue-purple

bands on the strips were specified as the lower detection limit of the assay. In order to determine the specificity, four different mixtures of the positive control were prepared as shown in table 2. After that, each mixture was subjected to the designed LiPA according to the above description. For further investigation of the probability of non-target genes detection in other organisms, the genome of negative control bacteria (*Klebsiella pneumonia* ATCC 7881, *E. coli* ATCC 25922, *Bacillus subtilis* ATCC 6051, *Staphylococcus aureus* ATCC 25923, *Enterococcus faecalis* ATCC 29212, *Streptococcus pneumoniae* ATCC 700669 and *Neisseria meningitidis* ATCC13060) was studied by the designed assay.

Artificially contaminated blood samples

Artificially infected blood samples were used in order to compare the in-house Line Probe Assay and SYBR Green Real-Time PCR concerning the detection of periodontal pathogens. For this purpose, a normal human EDTA-treated whole blood sample was utilized. The sample was distributed into thirty micro tubes (200 μ l/tube) which were subsequently spiked with the genomic DNA of the five periodontal pathogens as follows: The tubes were inoculated with 1 μ g/200 μ l final concentration of the genomic DNA of each pathogen. Moreover, ten blood samples (not spiked with DNA) were used as negative control. The artificially contaminated samples and the negative control samples were kept at 4°C for 96 hr. Then, forty samples were subjected to DNA extraction using EZ-10 spin Column Genomic DNA Kit (DNP™ kit (Bio Basic Inc., Canada). The DNA samples were examined by the in-house Line Probe Assay as mentioned above. Also, SYBR Green Real-time PCR was employed to detect the pathogens in the samples. ABI 7500 Fast Real-Time Amplification system (ABI, USA) and SYBR® Premix Ex Taq™ II (TAKARA, Japan) were utilized in all experiments. Each extracted DNA and the negative control were subjected to the SYBR Green Real Time PCR to detect each of the five periodontal pathogens, separately. Additionally, the purified genomic DNAs were used as positive control. The reaction tubes were prepared in a final volume of 20 μ l, containing 2 μ l of extracted genomic DNA as a template, 2 μ l of 2X master mix and 0.4 M of each of the forward and reverse primers (Table 3). The thermal

Table 3. Characteristics of the primer pairs utilized in the study

Bacterium	Target gene	Primer name	Position	Primer sequence	Reference
<i>A. actinomycetemcomitans</i>	<i>hbpA</i>	F-Aggrega- <i>hbpA</i>	2614 - 2634	AGACCCAATGCAAAAGTAACG	(7)
		R- Aggrega- <i>hbpA</i>	2774 - 2756	GCAGTTCTGGGCTGAATTG	
<i>P. gingivalis</i>	<i>fimA</i>	F- Porphy- <i>fimA</i>	296 - 313	ACAGCAGGAAGCCATCAA	Unpublished data
		R- Porphy- <i>fimA</i>	457 - 438	GCAGTCAGTTCAGTTGTCAA	
<i>P. intermedia</i>	<i>16S rRNA</i>	F- Pre 16	357-375	GAGGCAGCAGTGAGGAATA	Unpublished data
		R- Pre 16	637- 618	GCAAGGTAGATGTTGAGCAC	
<i>T. forsythia</i>	<i>16S rRNA</i>	F- Tan-16	469- 490	GCATGTACCTTGTGAATAAGCA	(7)
		R- Tan-16	718 -700	CTTCGCAATCGGAGTTCTG	
<i>T. denticola</i>	<i>16S rRNA</i>	F- Tre-16	266 - 284	CAAGGCAACGATGGGTATC	Unpublished data
		R- Tre-16	438 - 414	CTGCAAAAGAATTTACAACCTTTC	

cycling program was a 10 min activation of Taq polymerase at 95 °C by 40 cycles of PCR (denaturation at 95 °C for 15 s, annealing and extension at 60 °C for 1 min). Finally, to verify the correct product by its specific melting temperature (T_m), a melting curve analysis of the amplified DNA was performed at 54 to 95 °C and a temperature increase rate of 0.2 °C/sec was considered in this test.

Statistical analysis

The online Clinical Calculator 1 was used to calculate the clinical sensitivity and specificity of the LiPA and the SYBR Green Real-time PCR (<http://vassars-tats.net/clin1.html>). Clinical sensitivities of the assays were compared by McNemar's test using an online calculator (<https://graphpad.com/quickcalcs/mcNemar1/>).

Results

LiPA setting up

A 466 bp region from the *16s rRNA* gene of the five target bacteria was amplified and visualized on gel agarose. The PCR product lengths were the same regardless of the bacterial species (Figure 1). In preparing the positive control plasmids, the 466 bp PCR products were separately cloned in pTZ57R/T plasmid. The PCR and sequencing of the inserts confirmed the recombinant plasmids (pTZ-agg, pTZ-tan, pTZ-tre, pTZ-por and pTZ-pre). In the setup of the in-house LiPA, each target gene was minutely detected. In all strips, one universal probe was used to detect bacterial *16S rRNA* gene (Ampli control) and a conjugate control (Conj control) to confirm biotin and streptavidin reaction. In all runs, bands corresponding to Ampli control and Conj control showed blue-purple precipitate; however, in negative control cases (without DNA) only the band of Conj control was visualized. In addition, in each experiment, only the blue-purple precipitate related to the applied specific biotinylated product was produced (Figure 2).

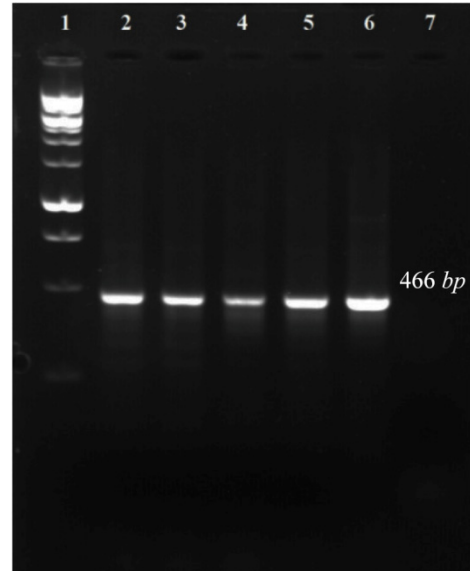


Figure 1. PCR amplification of 16s rRNA gene related to five target bacteria. 1: 100 bp DNA ladder, 2-6: negative control.

Analytical sensitivity and specificity results

Sensitivity results showed that the last dilution of the positive control plasmids (pTZ-agg, pTZ-tan, pTZ-tre, pTZ-por and pTZ-pre), generating a blue-purple precipitate on the strips, was a 10 fg dilution. Accordingly, the LOD of the test was determined 10 fg or 2760 copies of each of the five target genes. To determine the specificity, the in-house LiPA was performed on four different mixtures of the positive control plasmids. In each experiment, only signals corresponding to the specific biotinylated products were produced (Figure 3). Using the negative control bacterial genomes in the in-house LiPA, only bands pertaining to Ampli control and Conj control indicated the blue-purple precipitate on the strip.

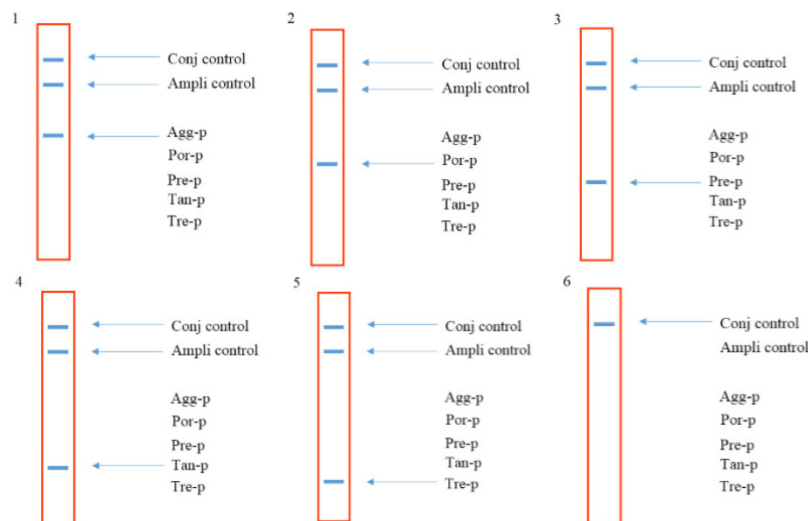


Figure 2. The schematic strips demonstrating the presence of the bacterial species based on the reaction between specific probes and corresponding biotinylated PCR product. 1: Aa, 2: Pg 3: Pi, 4: Tf, 5: Td and 6: negative control (without DNA).

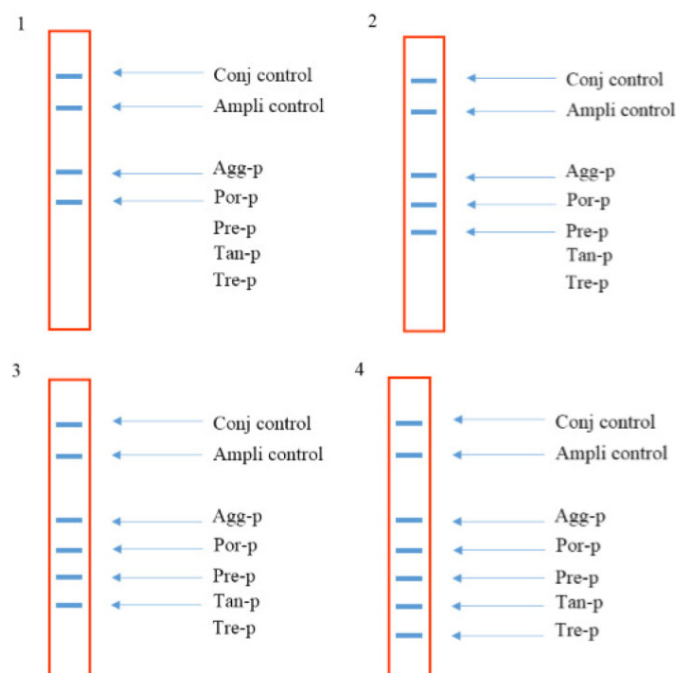


Figure 3. The schematic strips demonstrating the specificity of the in-house LiPA. 1) signals produced when using pTZ-agg and pTZ-por plasmids, 2) signals produced when using pTZ-agg, pTZ-por and pTZ-pre plasmids, 3) signals produced when using pTZ-agg, pTZ-por, pTZ-pre and pTZ-tan plasmids and 4) signals produced when using pTZ-agg, pTZ-por, pTZ-pre, pTZ-tan and pTZ-tre plasmids.

Results of artificially contaminated blood samples

The results of performing the LiPA on the DNA extracted from the spiked and non-spiked blood samples were inspected based on the relevant bands on the strips. In SYBR Green Real-time-PCR, a sample is considered positive when the amplification plot crosses the threshold value. The amplification plot of Aa contaminated samples exceeded the threshold baseline at a Ct value of 10-18, whereas the amplification plot of the negative control samples did not exceed the threshold baseline. Figures 4A, C, E, G, I show the Ct values of amplification plot related to the five periodontal pathogens. To ensure that the amplification plot obtained for the artificially contaminated sample was precise enough, Tm analyses of the assays were performed (Figures 4B, D, F, H, J). Table 4 illustrates the diagnostic sensitivity and specificity of the in-house LiPA and McNemar p-value when the SYBR Green Real-time PCR was used as gold standard. All the negative samples in the periodontal pathogen DNA (non-spiked samples) of the LiPA were also negative in SYBR Green Real-time PCR assay. Comparison of the results indicated no significant difference between the two methods (p-value=0.4795 for Aa, Pi and Td and 1 for Pg and Tf).

Discussion

In this study, 16S rRNA-based in-house LiPA was designed and applied so as to detect five major putative periodontal pathogens. 16S rRNA is an important housekeeping gene that shares a high degree of similar-

ity in different bacterial species. Using CLC Main Workbench software, the unique regions of the gene in each target bacterium were found and used as species-specific probe. Culture-based methods are technique-sensitive, time-consuming and expensive. In addition, they require a great amount of expertise and are difficult to perform, rendering such methods inappropriate for routine bacterial screening in the periodontal identification²⁹. Nowadays, the PCR method has become a powerful tool for the identification of various organisms, particularly those showing problems with regard to phenotypic methods.

Bacterial *16S rRNA* genes contain the interspersions of more- and less-conserved sequences; hence their application as a diagnostic gene in molecular detection is reasonable. The more distinct regions can be used to discriminate the bacterial taxa, while the conserved regions present universal sequences in annealing the PCR primers³⁰. Several studies have demonstrated the usefulness of 16S rRNA-based PCR detection of periodontal bacteria in subgingival samples. 16S rRNA PCR methods are more sensitive than culture-based approaches and less cross-reactive compared with DNA probe detection^{7,11,31,32}.

In this study, a pair of broad range universal bacterial primers was used²⁸ to amplify the *16S rRNA* gene of five target bacteria. The sequence alignment was used to identify a specific region of each *16S rRNA* gene, hence a candidate for specific probe. The preparation, hybridization, and development of the strips were subsequently performed.

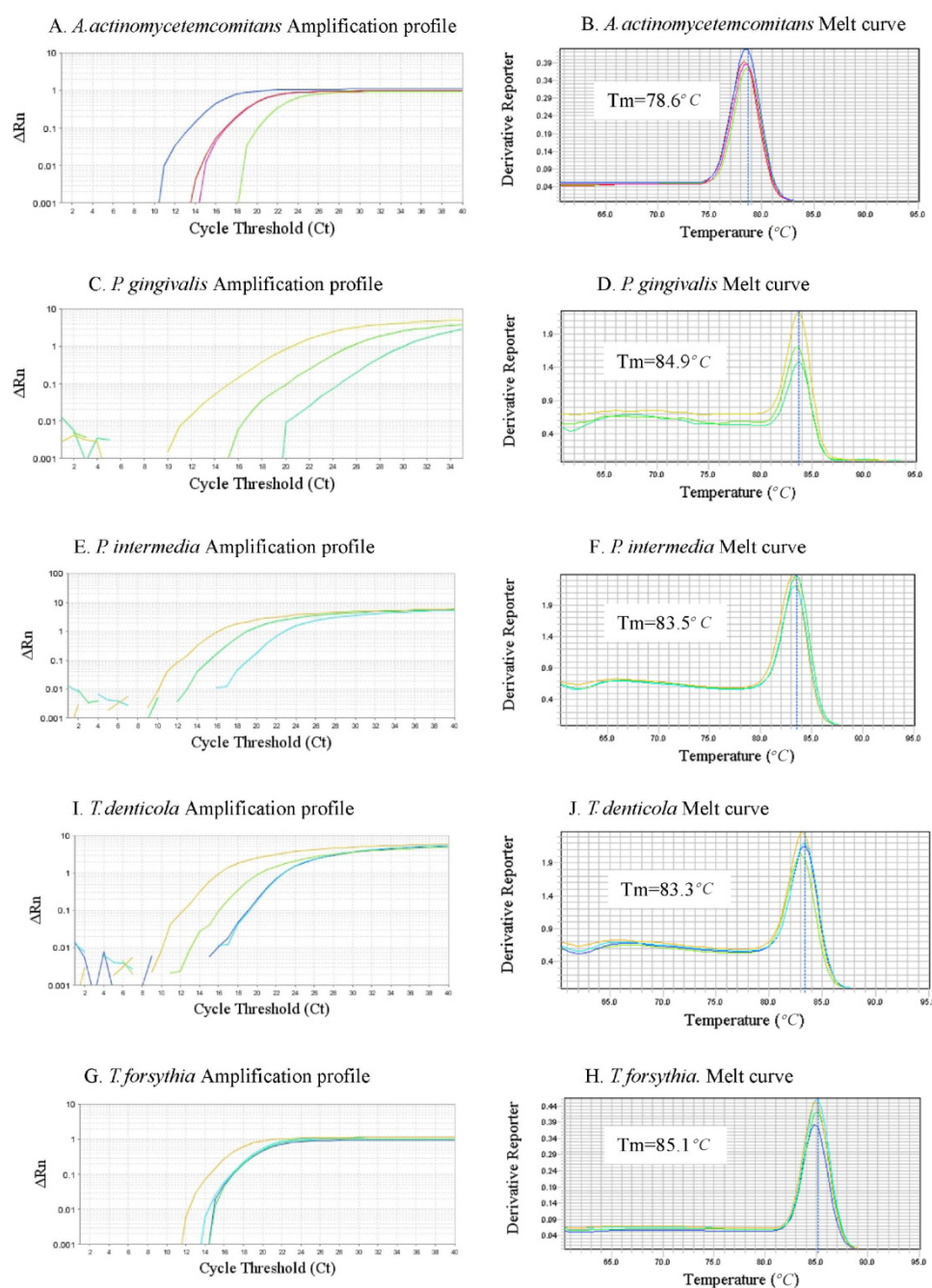


Figure 4. The amplification profiles and the corresponding melt curves of the periodontal pathogen, the melting temperature (T_m) of each amplicon is shown along with its melt curve.

In a previous study, to genotype Hepatitis C Virus (HCV) types and subtypes, the suitability of the workflow was shown, particularly the use of Biotin Chromogenic detection kit for the development of the strips³³. This is the first time that some of the most important periodontal pathogens are evaluated by an in-house line probe assay. With the in-house LiPA, the extracted DNA of artificially contaminated blood samples was evaluated and the results were compared with the SYBR Green Real-time PCR. It was concluded that there is concordance between results of the LiPA and

that of SYBR Green Real-time PCR as far as detecting the five periodontal pathogens is concerned. Aberle *et al* compared LiPA detecting drug-resistant Hepatitis B Virus (HBV) strains with sequence analysis³⁴. They obtained results concordant with sequence analysis with 48 of 56 serum samples from HBV-infected patients undergoing lamivudine therapy. Several studies have proposed LiPA as a screening test with low turnaround time, especially in low-resource regions^{35,36}. For example, Albert *et al* showed its usefulness for rapid screening of MDR-TB in Uganda³⁵. Melchers *et al*

Development and Comparison of In-house Line Probe Assay (LiPA) and SYBR Green Real-time PCR

Table 4. Comparison of SYBR Green Real-time PCR and in-house LiPA concerning the detection of DNA extracted from artificially contaminated blood samples

	SYBR Green Real- time PCR+	SYBR Green Real time PCR-	Total	Diagnostic sensitivity	Diagnostic specificity	McNemar p-value
<i>A. actinomycetemcomitans</i>						
LiPA+	27(67.5)	1(2.5)	28	96.4%	91.6 %	0.4795
LiPA-	1(2.5)	11(27.5)	12			
Total	28	12	40			
<i>P. intermedia</i>						
LiPA+	27(67.5)	1(2.5)	28	96.4%	91.6 %	0.4795
LiPA-	1(2.5)	11(27.5)	12			
Total	28	12	40			
<i>T. denticola</i>						
LiPA+	28(70)	1(2.5)	29	96.5%	90.9 %	0.4795
LiPA-	1(2.5)	10(25)	11			
Total	29	11	40			
<i>P. gingivalis</i>						
LiPA+	29(72.5)	0	29	96.6%	100 %	1
LiPA-	1(2.5)	10(25)	11			
Total	30	10	40			
<i>T. forsythia</i>						
LiPA+	29(72.5)	0	29	100%	100 %	1
LiPA-	0	11(27.5)	11			
Total	29	11	30			

applied LiPA to detect and genotype 16 different Human Papillomaviruses (HPV) types simultaneously, where it was shown that the method was sensitive, specific, simple, and fast. Also, they proposed HPV-LiPA to mass screen the cervical samples for routine analyses and patient management³⁷.

Conclusion

To screen periodontal pathogens, a simple, inexpensive and accurate method is required, so a reliable, sensitive, specific and economic detection assay was developed and applied in this study. LiPA provides the possibility of evaluating more species without the need for gel agarose electrophoresis step and can, therefore, be a useful candidate for research and diagnostic purposes of oral microbiology.

Acknowledgement

This study was conducted with the contributions of the members of the Research Council of the Islamic Azad University (Qom branch). The staff of microbiology laboratory is also sincerely appreciated.

References

- Hyvärinen K, Laitinen S, Paju S, Hakala A, Suominen-Taipale L, Skurnik M, et al. Detection and quantification of five major periodontal pathogens by single copy gene-based real-time PCR. *Innate Immun* 2009;15(4):195-204.
- Sheiham A, Netuveli GS. Periodontal diseases in Europe. *Periodontol* 2000 2002;29(1):104-121.
- Brown LJ, Löe H. Prevalence, extent, severity and progression of periodontal disease. *Periodontol* 2000 1993;2: 57-71.
- Chala S, Abouqal R, Abdallaoui F. Prevalence of apical periodontitis and factors associated with the periradicular status. *Acta odontol Scand* 2011;69(6):355-359.
- Slots J. The predominant cultivable microflora of advanced periodontitis. *Scand J Dent Res* 1977;85(2):114-121.
- Haffajee AD, Cugini MA, Tanner A, Pollack RP, Smith C, Kent RL Jr, et al. Subgingival microbiota in healthy, well-maintained elder and periodontitis subjects. *J Clin Periodontol* 1998;25(5):346-353.
- Soleimani M, Zolfaghari MR, Morovvati A. Development and comparison of conventional PCR and SYBR green real time PCR for detection of aggregatibacter actinomycetemcomitans and Tannerella forsythensis. *Jundishapur J Microbiol* 2013;6(8):e6757.
- Slots J, Ting M. Actinobacillus actinomycetemcomitans and porphyromonas gingivalis in human periodontal disease: occurrence and treatment. *Periodontol* 2000 1999; 20(1):82-121.
- Kozarov EV, Dorn BR, Shelburne CE, Dunn WA Jr, Progulske-Fox A. Human atherosclerotic plaque contains viable invasive actinobacillus actinomycetemcomitans and porphyromonas gingivalis. *Arterioscler Thromb Vasc Biol* 2005;25(3):e17-e8.
- Suzuki N, Yoshida A, Saito T, Kawada M, Nakano Y. Quantitative microbiological study of subgingival plaque by real-time PCR shows correlation between levels of tannerella forsythensis and fusobacterium spp. *J Clin Microbiol* 2004;42(5):2255-2257.
- Ashimoto A, Chen C, Bakker I, Slots J. Polymerase chain reaction detection of 8 putative periodontal pathogens in subgingival plaque of gingivitis and advanced periodontitis lesions. *Oral Microbiol Immunol* 1996;11 (4):266-273.

12. Maeda H, Fujimoto C, Haruki Y, Maeda T, Kokeguchi S, Petelin M, et al. Quantitative real-time PCR using TaqMan and SYBR Green for actinobacillus actinomycetemcomitans, porphyromonas gingivalis, prevotella intermedia, tetQ gene and total bacteria. FEMS Immunol Med Microbiol 2003;39(1):81-86.
13. Coffey J, Choudhry M, Shlossman M, Makin IRS, Singh VK. Multiplex real-time PCR detection and relative quantification of periodontal pathogens. Clin Exp Dent Res 2016;2(3):185-192.
14. Wen D, Zhang C. Universal multiplex PCR: a novel method of simultaneous amplification of multiple DNA fragments. Plant Methods 2012;8(1):32.
15. Wong ML, Medrano JF. Real-time PCR for mRNA quantitation. Biotechniques 2005;39(1):75-85.
16. Ling DI, Zwerling AA, Pai M. GenoType MTBDR assays for the diagnosis of multidrug-resistant tuberculosis: a meta-analysis. Eur Respir J 2008;32(5):1165-1174.
17. Abe C, Ogata H, Kawata K, Hiraga T, Takashima T, Suetake T. [Detection of rifampin-resistant Mycobacterium tuberculosis by line probe assay (LiPA)]. Kekkaku 2000;75(10):575-581. Japanese.
18. Naidoo P, du Toit E, Dunbar R, Lombard C, Caldwell J, Detjen A, et al. A comparison of multidrug-resistant tuberculosis treatment commencement times in MDRTB-Plus line probe assay and Xpert(R) MTB/RIF-based algorithms in a routine operational setting in Cape Town. PLoS One 2014;9(7):e103328.
19. Ritter C, Lucke K, Sirgel FA, Warren RW, van Helden PD, Bottger EC, et al. Evaluation of the AID TB resistance line probe assay for rapid detection of genetic alterations associated with drug resistance in Mycobacterium tuberculosis strains. J Clin Microbiol 2014;52(3):940-946.
20. Menton JF, Kearney K, Morgan JG. Development of a real-time RT-PCR and reverse line probe hybridisation assay for the routine detection and genotyping of noroviruses in Ireland. Virol J 2007;4:86.
21. Wang J, Bakkers J, Melchers WJ. [Genotyping of human papillomavirus genotypes in cervical scrapes by line probe assay]. Zhonghua Shi Yan He Lin Chuang Bing Du Xue Za zhi 1999;13(1):9-12. Chinese.
22. Noppornpanth S, Sablon E, De Nys K, Lien TX, Bro-uwer J, Van Brussel M, et al. Genotyping hepatitis C viruses from Southeast Asia by a novel line probe assay that simultaneously detects core and 5' untranslated regions. J Clin Microbiol 2006;44(11):3969-3974.
23. Andonov A, Chaudhary RK. Subtyping of hepatitis C virus isolates by a line probe assay using hybridization. J Clin Microbiol 1995;33(1):254-256.
24. Avó AP, Agua-Doce I, Andrade A, Pádua E. Hepatitis C virus subtyping based on sequencing of the C/E1 and NS5B genomic regions in comparison to a commercially available line probe assay. J Med Virol 2013;85(5):815-822.
25. Cai Q, Zhao Z, Liu Y, Shao X, Gao Z. Comparison of three different HCV genotyping methods: core, NS5B sequence analysis and line probe assay. Int J Mol Med 2013;31(2):347-352.
26. Comanor L, Elkin C, Leung K, Krajden M, Kronquist K, Nicolas K, et al. Successful HCV genotyping of previously failed and low viral load specimens using an HCV RNA qualitative assay based on transcription-mediated amplification in conjunction with the line probe assay. J Clin Virol 2003;28(1):14-26.
27. Liu CH, Liang CC, Liu CJ, Lin CL, Su TH, Yang HC, et al. Comparison of abbot realTime HCV genotype II with versant line probe assay 2.0 for Hepatitis C virus genotyping. J Clin Microbiol 2015;53(5):1754-1757.
28. Nadkarni MA, Martin FE, Jacques NA, Hunter N. Determination of bacterial load by real-time PCR using a broad-range (universal) probe and primers set. Microbiology 2002;148(Pt 1):257-266.
29. Baehni P, Guggenheim B. Potential of diagnostic microbiology for treatment and prognosis of dental caries and periodontal diseases. Crit Rev Oral Biol Med 1996;7(3):259-277.
30. Frank JA, Reich CI, Sharma S, Weisbaum JS, Wilson BA, Olsen GJ. Critical evaluation of two primers commonly used for amplification of bacterial 16S rRNA genes. Appl Environ Microbiol 2008;74(8):2461-2470.
31. Geevarghese A, Baskaradoss JK, Al Dosari A. Application of 16s rRNA in identifying oral microflora-a review of literature. World Appl Sci J 2012;17(10):1303-1307.
32. Hutter G, Schlagenhauf U, Valenza G, Horn M, Burge-meister S, Claus H, et al. Molecular analysis of bacteria in periodontitis: evaluation of clone libraries, novel phylogenotypes and putative pathogens. Microbiology 2003;149(Pt 1):67-75.
33. Majidzadeh-A K, Morovvati A, Soleimani M, Lange-roudi AG, Merat S, Jabbari H. Development and application of an In-house line probe assay for Hepatitis C virus genotyping. Hepat Mon 2013;13(5):e6767.
34. Aberle SW, Kletzmayer J, Watschinger B, Schmied B, Vetter N, Puchhammer-Stöckl E. Comparison of sequence analysis and the INNO-LiPA HBV DR line probe assay for detection of lamivudine-resistant hepatitis B virus strains in patients under various clinical conditions. J Clin Microbiol 2001;39(5):1972-1974.
35. Albert H, Bwanga F, Mukkada S, Nyesiga B, Ademun JP, Lukyamuzi G, et al. Rapid screening of MDR-TB using molecular line probe assay is feasible in Uganda. BMC Infect Dis 2010;10:41.
36. Barnard M, Gey van Pittius NC, van Helden PD, Bosman M, Coetzee G, Warren RM. The diagnostic performance of the genoType MTBDRplus version 2 line probe assay is equivalent to that of the Xpert MTB/RIF assay. J Clin Microbiol 2012;50(11):3712-3716.
37. Melchers WJ, Bakkers JM, Wang J, de Wilde PC, Bonstra H, Quint WG, et al. Short fragment polymerase chain reaction reverse hybridization line probe assay to detect and genotype a broad spectrum of human papillomavirus types. Clinical evaluation and follow-up. Am J Pathol 1999;155(5):1473-1478.