

A Multidisciplinary Study to Evaluate the Anti-quorum Sensing Ability of Phyto-compounds in *Ruellia patula* Jacq

Chemmutgil P¹, Lakshmi PTV^{1*}, and Annamalai A²

1. Centre for Bioinformatics, Pondicherry University, Pondicherry, India

2. PG and Research Department of Botany, Arignar Anna Government College, Thiruvalluvar University, Villupuram, Tamilnadu, India

Abstract

Background: *Staphylococcus aureus* (*S. aureus*) causing numerous diseases in humans, have become resistant to antibiotics, hence, urging the need for alternative medicines.

Methods: In this study, the Indian medicinal weed, *Ruellia patula* (*R. patula*) extracted and fractioned through column chromatography was subjected to antibacterial and anti-quorum sensing activity against *S. aureus* and Methicillin Resistant *Staphylococcus aureus* (MRSA).

Results: The obtained results confirmed fraction F44 to have significant effect as antimicrobial and anti-biofilm agent against both the micro-organism. Therefore, few of such highly active fractions were chemical finger printed using GC-MS and the compounds identified were further docked with DNA binding (LytTR) domain of *agrA*, which revealed that compounds identified from fraction were interactive to the protein.

Conclusion: *R. patula* is promising antimicrobial and anti-biofilm agent against *S. aureus* and MRSA.

Avicenna J Med Biotech 2019; 11(1): 48-58

Keywords: Antibiotics, DNA binding domain, *Methicillin resistant Staphylococcus aureus*, Quorum sensing, *Staphylococcus aureus*

Introduction

New antibiotics are obtained either by chemical synthesis¹ or developed from natural sources. According to WHO, 80% of the world population use plant extracts as medicine due to its inexpensive and lower side effects². Strong antimicrobial fractions derived from plants are reported to inhibit bacterial growth by mechanisms like membrane disruption, metabolism damage, anti-biofilm, bacterial capsule prevention and virulence attenuation¹. In this study, *Ruellia patula* (*R. patula*) Jacq (syn. *Dipteracanthus patulus* Nees) popularly called as kiranthi nayagam, chilanthinayagam, kayap-pachilai, Tutadi and sisodi belonging to Acanthaceae has been explored to identify its potential as the anti-biofilm agent. *R. patula* is distributed in tropical and sub-tropical regions of Africa, Arabia, Sri Lanka, Pakistan, Burma and India especially in Rajasthan, Haryana, Tamil Nadu, Western Ghats and Andhra Pradesh³. It is an erect, woody plant growing up to 50 cm and has been used as a medicine for a variety of problems including gonorrhea, syphilis, sore eyes, renal infection, cough, scalds, stomach ache, kidney stones, tumors, rheumatic complaints, dental problems, itches, insect

bites, paronychia, venereal diseases, for wound healing and as a cardiotonic and antiulcer agent. Juice of leaves acts as a sedative, roots as antipyretic agent, leaf paste as anti-inflammatory and flower and raw fruits as anti-diabetic agents⁴. Thus, based on these medicinal values, preliminary investigations on the selection of the most suitable solvents for better recovery of compounds have been explored in this research⁵.

In humans, gram positive bacteria *Staphylococcus aureus* (*S. aureus*) is responsible for a variety of infections including skin soft tissue infections⁶, and infective endocarditis, urinary tract infection, acute septic arthritis and hospital acquired infections like device-related infection⁷ especially under immune-suppression after organ transplantation and during chemotherapy¹ and bloodstream infections⁸. Since *S. aureus* is able to produce biofilms which restrict diffusion of antibiotic and results 20 to 1000 times more drug concentration in comparison to planktonic bacteria⁹, and is also antibiotic resistant¹⁰ and thereby decrease the number of patients up to 30%¹¹, it is important to focus more on alternative strategies to develop new antibiot-

* Corresponding author:
Lakshmi PTV, Ph.D., Centre for
Bioinformatics, Pondicherry
University, Pondicherry, India
Tel: +91 9486383094
E-mail:
lakanna@bicpu.edu.in;
lakshmiptv@yahoo.co.in
Received: 12 Jul 2017
Accepted: 31 Oct 2017

ics¹². Since pathogenicity or adhesion and protection against host defenses of *S. aureus* strongly depends on the virulence factors such as production of exotoxin, enzymes and cell wall associated proteins, which are reported to be regulated by the accessory gene regulator (agr) of regulatory locus¹³, and that its expression is reported to contribute to pathogenesis of biofilm associated infections in several models like murine subcutaneous abscesses, arthritis and rabbit endocarditis including invasion and apoptosis of epithelial cells¹⁴, agr protein was identified as the target in the present study. Agr system contains *agrB*, *agrD*, *agrC*, and *agrA* genes, of which, *agrB* and *agrD* genes get involved in the production of Auto Inducer peptide (AI), which binds to agrC (Transmembrane protein) to induce transcription by activating the agrA (response regulator)¹⁵. Thus, agrA up-regulates or represses the gene expression by binding the cognate DNA with DNA binding domain (also called LytTR domain), thereby regulates the expression of the gene involved in exotoxin synthesis¹⁵ and controls the production of virulence factor in *S. aureus*^{15,16}.

Moreover, LytTR domain is reported to be present in all response regulators of prokaryotes (approx. 2.7%) and observed in all *S. aureus* strains identified including Methicillin Resistant Staphylococcus aureus (MRSA) along with certain other pathogens such as *Clostridium* spp. and *Listeria* spp. However absent in human¹⁷ is of importance to us to be chosen as the target. Also, adequate number of subjects for the study is an important issue for conducting the research. Hence, it was hypothesized to evaluate the antimicrobial and quorum sensing activity of the fractions eluted through column chromatography against *S. aureus* and MRSA. Further, an attempt was made to identify the compounds present in those highly active fractions through GCMS and evaluate the lead molecules for interacting capacity with DNA binding (LytTR) domain of agrA.

Materials and Methods

Chemicals and instruments

Mueller Hinton broth and agar, Whatman filter paper No.1, analytical grade solvents such as Hexane, Chloroform, Ethyl acetate, Methanol, Dimethyl sulfoxide (DMSO) were all purchased from Himedia. Microplate analysis was done using Spectra Max M2e Microplate reader, Spinco Biotech, India. JEOLGCMATE II operating in EI mode at 70 eV (Agilent Technologies 6890N Network) was used for GC-MS analysis and spectrum was matched with NIST database. Softwares such as Marvin sketch, Autodock Vina, Autodock 4.2, Chimera, LigPlot+v.1.4, Graphp-ad prism were all used in this research.

Fractionation through column chromatography

The whole plants of *R. patula* were shade dried, coarsely powdered (500 g) and extracted through cold maceration method with 2 L of 80% methanol at 40 °C for 72 hr in orbital shaker until the residues became

pale colored. It was filtered using muslin cloth followed by whatman no.1 filter paper and condensed by rotary evaporator to obtain semisolid extract (48.76 g). Further, the semisolid extract was suspended in 500 ml of water and partitioned with 500 ml of Hexane (4.85 g). 20 g of concentrated water fraction was subjected to column chromatography while the remaining extracts were stored at 4 °C. The column was eluted through gradient solvent system, starting with 100% Hexane and Hexane combined with methanol in the ratio of 10:0; 9:1; 8:2 up to 5:5, followed by chloroform and ethyl acetate in the above mentioned combinations. Collected fractions were combined based on TLC profile, where approximately 60 different fractions were obtained and stored at 4 °C for further analysis¹⁸.

Maintenance of bacterial cultures

Clinical isolates of *S. aureus* and MRSA were collected from Department of Microbiology, Pondicherry University, Pondicherry and maintained in the Centre for Bioinformatics, Pondicherry University.

Determination of antibacterial activity

In order to evaluate the antibacterial efficiency of the fractions, disk diffusion assay was followed¹⁹ to determine the sensitivity of different fractions of *R. patula*, where both *S. aureus* and MRSA cultured on Mueller Hinton broth were incubated at 37±1 °C for overnight. Suspension with 1×10⁶ CFU/ml was swabbed on the solid Mueller Hinton agar medium in petri dish. Disks with 20 µl of different fractions each were placed on the swabbed agar plates and incubated at 37±1 °C for 24 hr. After incubation, the zones formed around the disks were measured and zone of inhibition was calculated. DMSO, which had no activity on bacteria, was used to dissolve fractions. The tests were conducted in triplicates.

Determination of minimum inhibitory concentration (MIC)

Based on the results of the disk diffusion assay, those fractions which showed higher antibacterial activity were further evaluated for MIC by broth dilution method²⁰, where, two fold dilutions of fraction in the range of 100 µg/ml to 12.8 mg/ml were prepared by dissolving in DMSO. Wells were filled with 20 µl of overnight culture (1×10⁶ CFU/ml) and 180 µl of Mueller Hinton broth followed by 100 µl of different concentrations of highly active fractions. DMSO alone was used as the negative control while, culture alone was maintained as the positive control and different concentrations of fractions with medium and without inoculums were used as blank and incubated for 18 hr at 37 °C for absorption at 600 nm. Triplicates were maintained and MIC was determined by comparing the absorbance of fraction with control and expressed in µg/ml.

Determination of synergistic activity

Based on the results of the antimicrobial activities of the eluted fractions, those which showed better activity were further examined to prove their action com-

parable to the antibiotic disks. Thus, disk diffusion method²¹ was proceeded to determine synergic effect of active fractions (F25, F44 and F53) combined with commercial antibiotics like tetracycline (30 µg/disk), Chloramphenicol (30 µg/disk), and vancomycin (30 µg/disk). 20 µl of active fractions (1 mg/ml) were impregnated and placed on Mueller Hinton Agar plate to incubate for 18 hr at 37°C. The synergistic activity of fraction with the combination of drugs was determined by measuring the zone of inhibition.

Performance of time killing assay

Based on the above observation, fraction 44 was further investigated for time killing assay²². Next, 1 ml of each *S. aureus* and MRSA respectively and 1 ml of highly active fraction F44 (MIC determined concentration) were added and incubated at 37°C from which at intervals of 0, 1, 2, 6, 12 and 24 hr, 200 µl of fraction treated culture was harvested and serially diluted (10⁻⁵) to spread on Mueller Hinton agar to determine the killing rate by colony count. Untreated culture was kept as the control and the number of colonies counted was expressed as CFU/ml.

Performance of anti-biofilm adhesion assay

Anti-biofilm adhesion activity of all 60 fractions against *S. aureus* and MRSA was carried out⁷ in a 96 well plate that was pre coated with 200 µl of fractions (1 mg/ml) and was dissolved in DMSO and allowed to dry. 200 µl of DMSO alone coated was considered as negative control while, non-coated wells were left as positive controls. Approximately, 1×10⁶ CFU/ml of 20 µl of *S. aureus* and MRSA was inoculated into 180 µl of Mueller Hinton broth wells. 200 µl of Mueller Hinton broth added in to fraction pre-coated well was used as the blank and the experiment was done in triplicate. The plate was incubated for 24 hr without shaking at 37°C and after 24 hr, the medium with loosely bound cells was removed and allowed to dry and washed three times with PBS. The remaining adhered cells were stained with 0.1% crystal violet for 30 min. Excess colorant was removed by rinsing with sterile distilled water and air dried. Further, 200 µl of 80% ethanol was added to the wells and OD was absorbed at 570 nm. The mean of the three replicates was calculated and the percentage of biofilm adhesion inhibition was calculated using the following formula.

$$\text{Inhibition \%} = \frac{\text{OD control} - \text{OD sample}}{\text{OD control}} \times 100$$

Pre-formed biofilm disturbance assay

To determine the disturbance capacity of fractions, Meng *et al*'s⁷ method was used. Accordingly, 20 µl of 1×10⁶ CFU/ml of culture was inoculated into 180 µl of MHB and incubated for 24 hr at 37°C. Loosely bound cells were removed by PBS buffer. Fractions dissolved in DMSO (1 mg/ml) were added to the preformed bio-

film in the wells and maintained DMSO alone was considered as the negative control while preformed biofilm alone as the positive control. Fractions dissolved in DMSO and medium alone were added to the clean wells and used as blank. The setup was incubated for 24 hr at 37°C without shaking. Wells were washed thrice with PBS and stained to calculate the inhibition percentage using the above formula.

GC-MS analysis of highly active column eluted fraction of *R. patula*

Chemical finger printing of highly active fraction was analyzed using JEOL GCMATE II GC-MS operated in EI mode at 70 eV (Agilent Technologies 6890N). In brief, 1 µl of fractions (0.1% w/v) was introduced into HP5 MS silica column (30 m x 0.25 mm). Injector temperature was 220°C and column temperature was fixed as 50-250°C at the rate of 10°C min⁻¹. High pure helium was used as the carrier gas. Physical and chemical properties were determined by comparing the obtained spectra against spectral library of NIST EI-MS database which is the in-house retention index library of organic volatile compounds²³. Percentage of compounds was calculated using the formula:

$$\text{Percent compd 1} = \frac{\text{Area of compd1}}{\text{Total area}} \times 100$$

In silico protein-ligand interaction studies

Compounds identified from the F44 of *R. patula* through GC-MS chemical finger printing were drawn using Marvin sketch and stored as pdb files. The target protein DNA binding (LytTR) domain of AgrA was downloaded from PDB (PDB ID: 4G4K) and optimized by removing hetero-atoms and water molecules. Further side chains and missing atoms were added along with hydrogen to obtain a complete and refined structure using whatif server. DNA binding sites were identified through literature survey²⁴. AutoDockVina²⁵ was used to import all the selected compounds while Auto Dock 4.2²⁶ tool was used to check the affinity of ligand towards the active site of selected protein. Protein was prepared by adding Kollman charges and polar hydrogens. Ligand was prepared by adding Gasteiger charges and detecting root. Grid was generated by selecting active site residues (Ser231 to Ile238) where side chains of active site residues were flexible to interact with the ligand molecule (which is rotatable and stretches for interacting with neighboring ligand molecule). 50 long runs of docking were performed with Lamarckian Genetic Algorithm. However, different poses of interaction were observed in docking where the best fitting pose was selected for further visualization and analyzation of docked structure using Chimera software. Non bonded interactions were obtained using LigPlot+ v.1.4²⁷.



Statistics analysis

All experiments were carried out in triplicates. Graph pad prism 6 was used to calculate mean and IC 50 value. T-test was performed to compare the mean values with the significance set at $p < 0.05$.

Results

Antibacterial activity of fractions against *S. aureus* and MRSA

Disk diffusion method was performed to ascertain the antimicrobial effect of fractions against *S. aureus* and MRSA, which showed different degrees of antibacterial zones ranging from 0.5 to 1.5 cm (Table 1). Fraction F25 was highly active against MRSA, but no activity was observed in *S. aureus*. Unlike that, F53 was highly active against *S. aureus* while no activity was detected against MRSA. Interestingly, F44 was active against both *S. aureus* and MRSA. Generally, *S. aureus* is resistant to a number of antibiotics and pro-

duces enterotoxins²⁸. Presence of phytochemicals like fatty acids, phenols, flavonoids, alkaloids, terpenes and steroids reported in *R. patula* has proved their bioactive and antimicrobial activity²⁹. Fatty acids are considered as effective broad spectrum and non-specific antibiotics, which disrupt the oxidative phosphorylation and electron transport and also result in nutrient uptake impairment, thus inducing bacterial cell lysis³⁰. Therefore, different forms of fatty acids present in the extracts and polyphenolic compounds may play an important role as therapeutic agents and can attribute to antibacterial activity by different mechanisms like metabolism inhibition, membrane damage and inhibition of synthesis of cell wall, cell membrane and nucleic acid².

Terpenes like thymol, linalool and menthol have been reported to inhibit bacteria via membrane lipid bilayer penetration through fatty acyl chains and

Anti-quorum Efficiency of *R. patula*

Table 1. Determination of antimicrobial and anti-quorum sensing ability of *R. patula* fractions eluted by Column chromatography

S. No	Fractions	<i>S. aureus</i>			MRSA		
		Zone of inhibition (cm)	Anti-adhesion (%)	Disturbance (%)	Zone of inhibition (cm)	Anti-adhesion (%)	Disturbance (%)
1	F1	-	-	-	-	-	-
2	F2	-	-	-	-	-	-
3	F3	-	-	-	1.1	88.76	24.59
4	F4	-	-	-	-	-	-
5	F5	-	-	-	-	-	-
6	F6	-	-	-	-	-	-
7	F7	-	-	-	-	-	-
8	F8	-	-	-	-	-	-
9	F9	-	-	-	-	-	-
10	F10	-	-	-	-	-	-
11	F11	-	-	-	-	-	-
12	F12	-	-	-	1	85.61	70.6
13	F13	-	-	-	1	79.14	84.44 ^{##}
14	F14	-	-	-	-	-	-
15	F15	-	-	-	-	-	-
16	F16	-	-	-	-	-	-
17	F17	-	-	-	-	-	-
18	F18	-	-	-	1	-	-
19	F19	-	-	-	-	-	-
20	F20	-	-	-	-	-	-
21	F21	-	-	-	1	83.54	-
22	F22	-	-	-	-	-	-
23	F23	-	-	-	-	-	-
24	F24	-	-	-	-	-	-
25	F25	-	-	-	1.5 ^{**}	82.29	67.58
26	F26	-	-	-	-	-	-
27	F27	-	-	-	-	-	-
28	F28	-	-	-	1.4	82.08	27.01
29	F29	-	-	-	-	-	-
30	F30	-	-	-	-	-	-
31	F31	-	-	-	-	-	-
32	F32	-	-	-	1	82.6	-
33	F33	-	-	-	-	-	-
34	F34	-	-	-	-	-	-
35	F35	-	-	-	1.3	86.46	75.82
36	F36	-	-	-	-	-	-
37	F37	-	-	-	-	-	-
38	F38	1.3	80.6	60.7	-	-	-
39	F39	-	-	-	1.4	89.17	59.41
40	F40	1.4	90.06 [@]	77.2	-	-	-
41	F41	-	-	-	-	-	-
42	F42	-	-	-	-	-	-
43	F43	1.4	84.02	81.5 [#]	-	-	-
44	F44 ^{\$}	1.2	89.9	51.6	1.4	86.8	72.1
45	F45	-	-	-	-	-	-
46	F46	-	-	-	-	-	-
47	F47	-	-	-	-	-	-
48	F48	-	-	-	-	-	-
49	F49	-	-	-	-	-	-
50	F50	1.3	77.2	66.88	-	-	-
51	F51	-	-	-	-	-	-
52	F52	-	-	-	-	-	-
53	F53	1.5 [*]	64.8	48.93	-	-	-
54	F54	1.1	75.6	75.61	-	-	-
55	F55	-	-	-	1	95.2 ^{@@}	61.06
56	F56	-	-	-	-	-	-
57	F57	-	-	-	-	-	-
58	F58	-	-	-	1	87.32	73.47
59	F59	-	-	-	-	-	-
60	F60	-	-	-	-	-	-

* Highest inhibition zone for *S. aureus*. ** Highest inhibition zone for MRSA. @ Highest adhesion inhibition percentage of *S. aureus*. @@ Highest adhesion inhibition percentage of MRSA. # Highest biofilm disturbance percentage of *S. aureus*. ## Highest biofilm disturbance percentage of MRSA. \$ Fraction having all the activity against both *S. aureus* and MRSA.

disrupting lipid packing thus altering cell membrane fluidity²³. Flavonoids have been reported as antimicrobial agents, which by inhibiting the nucleic acid

synthesis damage cytoplasmic membrane and membrane functions by inhibiting the formation of D-alanine-Dalanine². Moreover, the antibacterial activity

of fraction could be correlated with the amount of phenolic and flavonoid content present in the fraction and this perhaps may not be due to the single compound, but attributed to the combinations of interacting compounds^{23,31}. Thus, the antibacterial activity of the fractions could be due to the presence of bioactive compounds such as terpenes, phenols and flavonoids which can act as antibacterial agents by any of the above mechanism supporting the fact that plant fractions could be used as antibacterial agent.

MIC of highly active fraction for respective organisms

MIC is the lowest concentration needed for complete inhibition of selected organism. In disk diffusion assay, the antibacterial activity depends on the diffusion nature of fractions and polarity of solvents used for fractionation which probably may contribute to higher activity at lower concentration or lower activity at higher concentration^{32,33}. Thus, to determine the IC₅₀ value, highly active fractions (F25, F53 and F44) were further subjected to MIC determination against both *S. aureus* and MRSA using two fold broth dilution method. MIC value of F25 was determined as 1.219 mg/ml against MRSA while, 3.147 mg/ml was recorded for F53 against *S. aureus*. In case of F44, ranges from 0 to 78.27±1.48% were obtained against *S. aureus*, while 0 to 82.05±1.7% of inhibition was obtained for MRSA at respective concentrations. The results suggested F44 to be effective against both *S. aureus* and MRSA even at lower concentrations of 1.922 mg/ml and 1009 µg/ml, respectively (Figure 1). MRSA had lower IC₅₀ value than *S. aureus* which shows that this fraction can inhibit MRSA effectively than *S. aureus*.

Synergistic effect of highly active fractions with the combination of commercial drugs against selected organisms

Following the effect of antibacterial activity (against both MRSA and *S. aureus*) exhibited by the F44 (unlike F23 and F53, which acted against any one pathogen), all three were subjected to synergistic activity

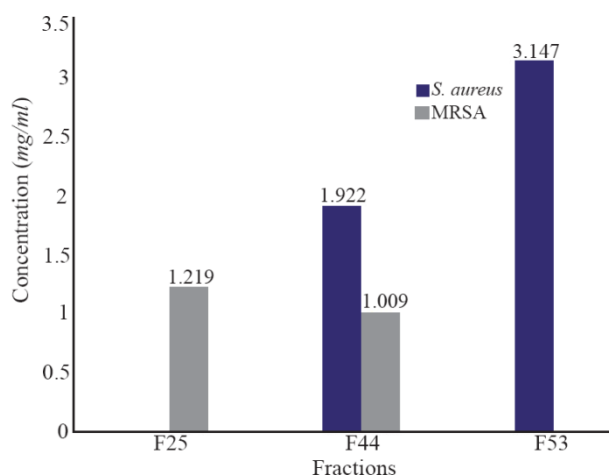


Figure 1. Determination of Minimum Inhibitory Concentration of highly active fractions of *R. patula*. The role of fractions indicate F44 to be active against both the selected strains of *S. aureus*.

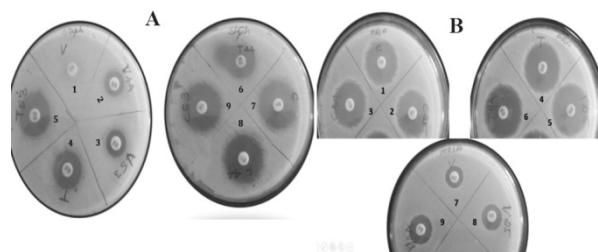


Figure 2. Determination Synergistic effect of highly active fractions with the combination of commercial drugs against *S. aureus* and MRSA. A) Synergistic effect of fractions on *S. aureus*; (1) Vancomycin alone; (2) Vancomycin with the combination of fraction F44; (3) Vancomycin with the combination of fraction F53; (4) Tetracycline alone; (5) Tetracycline with the combination of fraction F53; (6) Tetracycline with the combination of fraction F44; (7) Chloramphenicol alone; (8) Chloramphenicol with the combination of fraction F44; and (9) Chloramphenicol with the combination of fraction F44. B) Synergistic effect of fractions on MRSA; (1) Chloramphenicol alone; (2) Chloramphenicol with the combination of fraction F25; (3) Chloramphenicol with the combination of fraction F44; (4) Tetracycline alone; (5) Tetracycline with the combination of fraction F25; (6) Tetracycline with the combination of fraction F44; (7) Vancomycin alone; (8) Vancomycin with the combination of fraction F25; and (9) Vancomycin with the combination of fraction F44.

with commercial drugs such as chloramphenicol, vancomycin and tetracycline available as disks (each 30 µg/disk). Interestingly, the synergistic effect was not observed when the fractions were combined with chloramphenicol against both organisms. However, F53 alone when combined with tetracycline, synergistically reacted against *S. aureus*. The reason for tetracycline effectiveness and chloramphenicol failure may be due to the fact that, chloramphenicol as a broad spectrum bacteriostatic antibiotic, when combined with other antibiotics interfered with their action making them also inactive. This is perhaps evidenced by studies of Aqil *et al*³⁴, Beoni *et al*³⁵, Klastersky and Husson³⁶, and Jawetz *et al*³⁷ according to whom, synergistic interaction of chloramphenicol by the selected plant extracts and antibiotics against *S. aureus* actually failed, although it is a broad spectrum antibiotic which targets the cells protein synthesis but it still nullifies the effect by interfering with the other combined antibiotic.

Thus, accordingly, in our study too, the selected organism treated with chloramphenicol and fractions simultaneously failed due to the interference of chloramphenicol on activity of our fractions. However, vancomycin showed synergistic effect with the combination of fractions, but when vancomycin alone was tested, there was no activity detected. Interesting results obtained from this test showed that the clinically isolated *S. aureus* was vancomycin resistant (Figures 2A and 2B) thus suggesting the fact that compounds present in the fractions with good activity could be identified as hit for Vancomycin-Resistant Staphylococcus Aureus (VRSA) infections along with MRSA infections. Hence, it necessitated the purification of phytochemicals present in this plant, which further could be explored to develop novel antibacterial compounds for the treat-

ment of virulent and resistance strains of *Staphylococcus*.

Time killing assay of highly active fraction F44 against *S. aureus* and MRSA

Substances which kill or eliminate bacteria at the shortest time are considered bactericidal²⁹ whereas the ones that inhibit the bacterial growth are bacteriostatic³⁸. Accordingly, the shortest time taken by fraction F44 to kill or eliminate bacteria was determined by Time Killing assay (Figure 3), where MRSA colonies were reduced to 52.45% after one hr of contact to MIC concentration and further decreased to 39.34, 22.95 and 11.47% with increasing contact time of 6, 12 and 24 hr, respectively thus indicating the fact that the time exposed to fraction was directly proportional to the defense mediated by the fraction. However, this phenomenon was done to a greater extent (reduced to 90.32 in one hr and 77.41, 45.16 and 22.58% by the 6th, 12th and 24th hr respectively) in *S. aureus* indicating its suitability and favorability against those strains which are resistance to vancomycin, too (VRSA). Although based on the results, it could be observed that the selected fraction at determined concentration could not completely kill the organism at the given time (24 hr), it still had the ability to control the colony formation when compared to the control, suggesting the fact that fractions possess slow bactericidal activity against selected strains³⁹.

Anti-biofilm Assay

The research focused on plant derived alternative medicine against bacteria was extended to control biofilm formation and was performed in two ways.

Biofilm adhesion inhibition assay

Crystal violet assay was performed and different degrees of anti-adhesion effect by the fractions of *R. patula* were indicated to inhibit biofilm adhesion (Table 1). Twelve fractions showed activity towards MRSA (79.14-95.2%) and seven fractions had activity towards *S. aureus* (64.8-90.06%) revealing the effectiveness of

fractions more against MRSA than *S. aureus*. However, highest inhibition percentage (95.2%) was obtained by F55 against MRSA and the lowest was obtained by F53 (64.8%) against *S. aureus* whereas in antibacterial activity F53 showed high inhibitory activity. Interestingly, F44 exhibited activity against both *S. aureus* (89.9%) and MRSA (86.8%) because as evidenced phytochemicals present in that fraction could have been responsible for inhibiting the biofilm adhesion by altering cell surface and or by inhibiting cell interactions to substance. Moreover, adhesion to the surface is the primary step to form biofilm and it depends on the characteristics of bacteria as influenced by the physicochemical properties of surface and environment². Thus, accordingly, different degrees of inhibition in our study could be revealed by the presence of flavonols of other phytoconstituents in the fractions attributed to anti-adhesion effect using different mechanisms.

Disturbance assay of preformed biofilm

Disturbance effect of fractions on formed biofilm was evaluated which showed that fractions had different degrees of disturbance effect on *S. aureus* (48.93-81.5%) and MRSA (24.59-84.44%) (Table 1). Following the result of anti-adhesion assay, here too, F44 disturbed the biofilms of both *S. aureus* (51.6%) and MRSA (72.1%). It is difficult to control the formed biofilm ones and therefore they need to be inhibited at the time of biofilm initiation²³. Thus, the results of our study too consequently produced less disturbance than anti-adhesion. Although the mechanism of anti-biofilm is unknown, it is predicted that formed biofilm might be disturbed by the compounds present in the fractions by disturbing the process of adhesion through deactivating the enzymes responsible for surface attachment or cell-cell communication mediating signaling for cross-talks. Thus, accordingly, the formation of biofilm by *S. aureus* was inhibited by terpenes²³. Moreover, flavonoids such as apigenin, quercetin and myricetin were also reported to exhibit strong biofilm inhibition activity and exhibit enzyme modulation which is responsible for adherence of surface protein to the host cell and also responsible for virulence factors in surface protein². Hence, strong anti-biofilm activity in the present study could also be indicated by the adequate quantity of such compounds present in the fractions tested and therefore proceeded further to examine the compound identities.

GC-MS analysis

Fraction F44 was subjected to GC-MS for chemical finger printing analysis and nine compounds were identified (Figure 4). N-Hexadecanoic acid (Palmitic acid) was identified as the third compound and remarkably present in high concentration of 86.07% at RT 17.9 followed by Morin at RT 18.97 with 8.80%, while Oleic acid (RT 18.82), 4,8,12,16-Tetramethyl-heptadecan-4-olide (RT 21.2), 2-Cyclohexen-3,6-diol-1-one, 2-tetradecanoyl (RT 22.43) and Hexadecanoic acid, 2-bromo (23.63) were eluted with minimum concentra

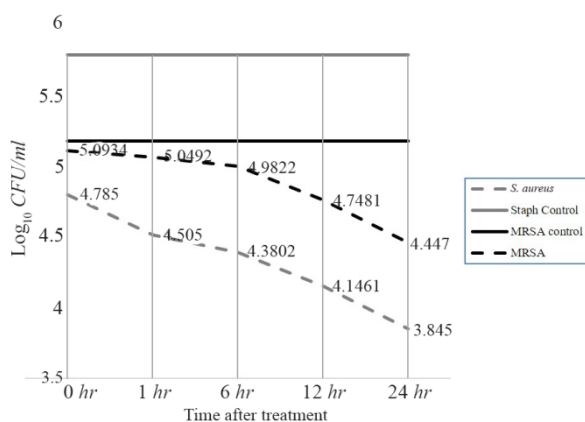


Figure 3. Time killing assay of fraction F44 against *S. aureus* and MRSA. The values represent the decrease in the growth to be directly proportional to the time.

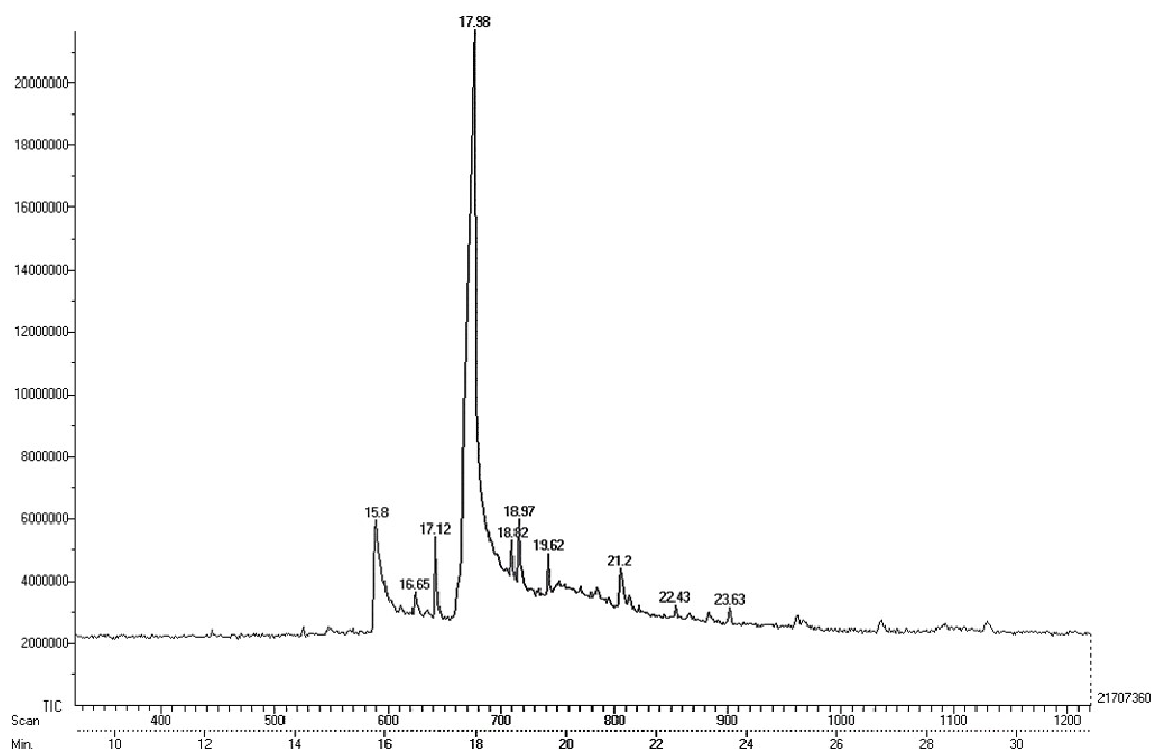


Figure 4. GC Chromatogram of highly active fraction F44. Finger print of the column eluted fraction44 demonstrates the peak area representing the separation of nine different compounds.

Table 2. Chemical finger prints of Fraction F44 by GC-MS

S. No	RT	Chemical name	Molecular formula	Molecular weight	Peak area %
1	15.8	Flavone	C ₁₅ H ₁₀ O ₂	222.239	2.492
2	17.12	4',5,7-Trihydroxy isoflavone	C ₁₅ H ₁₀ O ₅	270.237	2.160
3	17.98	n-Hexadecanoic acid *	C ₁₆ H ₃₂ O ₂	256.424	86.07
4	18.82	Oleic acid	C ₁₈ H ₃₄ O ₂	282.461	0.058
5	18.97	Morin	C ₁₅ H ₁₀ O ₇	302.236	8.809
6	19.62	Z-5,17-Octadecadien-1-ol acetate	C ₂₀ H ₃₆ O ₂	308.499	0.129
7	21.2	4,8,12,16-Tetramethylheptadecan-4-olide	C ₂₁ H ₄₀ O ₂	324.541	0.160
8	22.43	2-Cyclohexen-3,6-diol-1-one, 2-tetradecanoyl	C ₂₀ H ₃₄ O ₄	338.488	0.044
9	23.63	Hexadecanoic acid, 2-bromo	C ₁₆ H ₃₁ BrO ₂	335.326	0.070

* Compound present in highest concentration.

tion of below 1% (Table 2). Identification of these compounds in this fraction was helpful in understanding the mechanism of action of antibacterial and anti-biofilm activity. Anti MRSA activity achieved by flavanone is beneficial to the structure of A ring (5,7-dihydroxylation) and B ring (2',4'- or 2',6'-dihydroxylation), which is essential for significant activity⁴⁰. Thus, flavanone present in the fraction might be responsible for the antibacterial activity of MRSA. Genistein possesses exotoxin inhibition properties along with antibacterial activity⁴¹, which might be one of the reasons for the potential activity of the fraction. Ohta *et al*⁴² reported that palmitic acid had no antibacterial activity against *S. aureus* and MRSA, while Huang *et*

*al*⁴³ reported that, long chain, medium chain and short chain fatty acids contribute to inhibition of biofilm and thereafter evolution and emergence of new species could be restricted. Thus, the presence of fatty acids might support biofilm activity of the fraction. Moreover, poly phenolic compounds possess anti-biofilm activity either by inhibition or by anti-adhesion. The poly phenolic compounds like flavone, 4', 5,7-Trihydroxyisoflavone and Morin might be responsible for anti-biofilm activity of the fraction⁴⁴.

Molecular docking studies

AgrA gene plays a significant role in quorum sensing and survival of *S. aureus*. Nine different compounds (Table 2) identified through GC-MS for the

Table 3. Determination of binding energies of identified compounds with DNA binding (LytTR) domain of agrA using Auto Dock vina

S. No	Ligand	Binding energy (kcal/mol)
1	Flavone	-6
2	4',5,7-Trihydroxy_isoflavone	-6.8
3	n-Hexadecanoicacid	-3.6
4	Oleic acid	-3.8
5	Morin*	-7.1
6	Z-5,17-Octadecadien-1-ol_acetate	-4.1
7	4,8,12,16-Tetramethylheptadecan-4-olide	-4
8	2-Cyclohexen-3,6-diol-1-one, 2-tetradecanoyl	-4.3
9	Hexadecanoicacid, 2-bromo	-4.1

*Protein –compound complex, which had best binding energy.

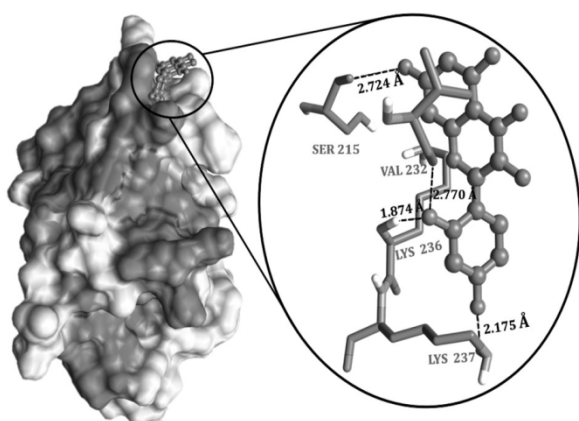


Figure 5. Docking interaction of Morin with DNA binding (LytTR) domain of AgrA. The interaction pattern highlights the efficiency of morin inhibiting the interactive residues at the target site.

fraction F44 were drawn through Marvin sketch and those files were subjected to AutodockVina interaction software, which accepts multiple compounds to understand their interaction ability on the single target protein. Thus, accordingly, nine different compounds subjected at the active site (Ser231 to Ile238) on the DNA binding domain of agrA protein, revealed that all compounds interact with the protein as evidenced through the calculated binding energies, which ranged from -3.6 kcal/mol to -7.1 kcal/mol. However, in comparison with the study of Leonard *et al*²⁴, the apo form of the protein interacted with the tested compounds and exhibited the binding energies between -3.5 kcal/mol to -4.1 kcal/mol for different compounds such as 4-phenoxyphenol, 9 H-xanthene-9-carboxylic acid, 2-(4-methylphenyl)-1,3-thiazole-4-carboxylic acid, *etc.*

Discussion

Thus, based on the above comparison, the compounds under investigation showed better occupancy; however, Morin (C₁₅H₁₀O₇) exhibited good interaction pattern with less binding energy of -7.1 kcal/mol, followed by genistein with -6.8 kcal/mol and flavone with -6 kcal/mol energy, respectively (Table 3). Therefore, Morin was proceeded to have a better insight of

molecular interactions using AutoDock 4 tool. The Grid box was generated considering the conserved regions between Ser231 to Ile238 which form a shallow groove for the DNA binding site and exhibit 100% identity among 211 *Staphylococcal* strains of Uniprot database²⁴. Morin interacted with the DNA binding region where it formed four hydrogen bonds (Table 4 and Figure 5). Both Val232-O and Lys236-NH exhibited hydrogen bond with O₂' of hydroxyphenyl moiety, while Lys237-H interacted through O₄' of hydroxyphenyl moiety and Ser215 bonded with Chromen-4-one moiety of Morin, respectively. Additionally, the interactions were stabilized by electrostatic (with Lys236 and Arg233) and hydrophobic interactions (with Val232 and Val235) with the domain region of agrA. Thus, from the interaction pattern, it is clear that the highly active residues, which are also considered as key residues involved in regulating the DNA binding activity at the active pocket, were actually engaged in bonding with our ligand-Morin, proving its ability to inhibit the activity of agrA and therefore mimic the DNA, which thereby may suppress the upregulation of virulence factor genes. However, the better binding energies compared to Leonard PG *et al*²⁴, may also be attributed to another important catalytic active polar residue SER215, which formed hydrogen bond through O7 of Chromen-4-one moiety of Morin, as this was not reported in earlier studies of Leonard *et al*²⁴ that may have influenced the formation of binding energies and could be responsible for the stronger interaction as evidenced through the bond lengths (Table 4). Hence, considering the fact that interaction of Morin at binding site could quench the quorum sensing ability and therefore disturb the cell wall synthesis and destruct the growth of the organism (unaccompanied data), if proven through wet lab studies, it may be applied for all 211 strains of *Staphylococcus* and other pathogens possessing LytTR domain in agrA proteins.

Conclusion

The work uncovered the importance of fractions of *R. patula* by means of anti-bacterial and anti-biofilm activity, thereby supported the medicinal use of this

Table 4. Hydrogen bond interactions of morin with DNA binding (LytTR) domain of AgrA

S. No	Residues	Atom of residues	Ligand atoms	Distance (Å)
1	LYS237	H ₂	O ₄ '	2.175
2	SER215	O	O ₇	2.724
3	VAL232	O	O ₂ '	2.770
4	LYS236	NH	O ₂ '	1.874

plant in siddha medicine. Fraction F44 was positive for all activities against both VRSA and MRSA. Chemical finger printing of F44 added valuable information to the anti-bacterial activity of that fraction. Hence, the phytochemicals present in *R. patula* can be used as anti-biotic against both VRSA and MRSA. Molecular docking of Morin with DNA binding domain of agrA revealed it as the potential agent for quorum sensing thereby enabling it to be used against all staphylococcus bacteria including MRSA and other pathogens. However, further research on the purification of this compound will enable to elucidate the mechanism of natural process of Morin on biofilm inhibition, which is also basically suggested as the futuristic study.

Acknowledgement

Authors thank Dr. Joseph Selvin, co-ordinator, Department of Microbiology, Pondicherry University for providing the cultures and lab facilities.

Conflict of Interest

The authors declare no conflict of interest.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

References

- Zbikowska B, Franciczek R, Sowa A, Polukord G, Krzyzanowska B, Sroka Z. Antimicrobial and antiradical activity of extracts obtained from leaves of five species of the genus *bergenia*: identification of antimicrobial compounds. *Microb Drug Resist* 2017;23(6):771-780.
- Nostro A, Guerrini A, Marino A, Tacchini M, Di Giulio M, Grandini A, et al. In vitro activity of plant extracts against biofilm-producing food-related bacteria. *Int J Food Microbiol* 2016;238:33-39.
- Yadav S, Arya V, Kumar S, Yadav JP. Anti-inflammatory activity of root, leaves and stem of *Dipteracanthus patulus* (Jacq.) Nees (Acanthaceae). *Asian Pac J Trop Biomed* 2012;2(1):S187-S191.
- Ramadevi S, Kaleeswaran B, Natarajan P. Phytochemicals analysis and antimicrobial activity of *Ruellia patula* L. against pathogenic microorganisms. *South Indian J Biol Sci* 2016;2(2):306-313.
- Lakshmi P. Exploring the Phyto-Constituents of *Ruellia patula* (Acanthaceae) as Antibacterial Agent. *EC Microbiol* 2017;7:133-148.
- Lacey KA, Geoghegan JA, McLoughlin RM. The role of *Staphylococcus aureus* virulence factors in skin infection and their potential as vaccine antigens. *Pathogens* 2016;5(1). pii: E22.
- Meng Y, Hou X, Lei J, Chen M, Cong S, Zhang Y, et al. Multi-functional liposomes enhancing target and antibacterial immunity for antimicrobial and anti-biofilm against methicillin-resistant *staphylococcus aureus*. *Pharm Res* 2016;33(3):763-775.
- Thomer L, Schneewind O, Missiakas D. Pathogenesis of *Staphylococcus aureus* bloodstream infections. *Annu Rev Pathol* 2016;11:343-364.
- Kashef N, Akbarizare M, Razzaghi MR. In vitro activity of Linezolid in combination with photodynamic inactivation against *staphylococcus aureus* biofilms. *Avicenna J Med Biotechnol* 2017;9(1):44-48.
- Vuong C, Yeh AJ, Cheung GY, Otto M. Investigational drugs to treat methicillin-resistant *Staphylococcus aureus*. *Expert Opin Investig Drugs* 2016;25(1):73-93.
- Van Hal SJ, Jensen SO, Vaska VL, Espedido BA, Paterson DL, Gosbell IB. Predictors of mortality in *Staphylococcus aureus* bacteremia. *Clin Microbiol Rev* 2012;25(2):362-386.
- den Reijer PM, Haisma EM, Lemmens-den Toom NA, Willemse J, Koning RI, Demmers JA, et al. Detection of alpha-toxin and other virulence factors in biofilms of *Staphylococcus aureus* on polystyrene and a human epidermal model. *PloS One* 2016;11(1):e0145722.
- Lina G, Jarraud S, Ji G, Greenland T, Pedraza A, Etienne J, et al. Transmembrane topology and histidine protein kinase activity of AgrC, the agr signal receptor in *Staphylococcus aureus*. *Mol Microbiol* 1998;28(3):655-662.
- Yarwood JM, Schlievert PM. Quorum sensing in *Staphylococcus aureus* infections. *J Clin Invest* 2003;112(11):1620-1625.
- Kong KF, Vuong C, Otto M. *Staphylococcus aureus* quorum sensing in biofilm formation and infection. *Int J Med Microbiol* 2006;296(2-3):133-139.
- Rajasree K, Fasim A, Gopal B. Conformational features of the *Staphylococcus aureus* AgrA-promoter interactions rationalize quorum-sensing triggered gene expression. *Biochem Biophys Rep* 2016;6:124-134.
- Galperin MY. Telling bacteria: do not LytTR. *Structure* 2008;16(5):657-659.
- Komape NP, Bagla VP, Kabongo-Kayoka P, Masoko P. Anti-mycobacteria potential and synergistic effects of combined crude extracts of selected medicinal plants used by Bapedi traditional healers to treat tuberculosis related symptoms in Limpopo Province, South Africa. *BMC Complement Altern Med* 2017;17(1):128.
- Zhang WM, Wang W, Zhang JJ, Wang ZR, Wang Y, Hao WJ, et al. Antibacterial constituents of hainan morinda citrifolia (Noni) Leaves. *J Food Sci* 2016;81(5):M1192-1196.
- Rosas-Burgos EC, Burgos-Hernandez A, Noguera-Artiaga L, Kacaniová M, Hernandez-Garcia F, Cardenas-Lopez JL, et al. Antimicrobial activity of pomegranate

- peel extracts as affected by cultivar. *J Sci Food Agric* 2017;97(3):802-810.
21. Su PW, Yang CH, Yang JF, Su PY, Chuang LY. Antibacterial activities and antibacterial mechanism of polygonum cuspidatum extracts against nosocomial drug-resistant pathogens. *Molecules* 2015;20(6):11119-11130.
 22. Gorle AK, Li X, Primrose S, Li F, Feterl M, Kinobe RT, et al. Oligonuclear polypyridylruthenium (II) complexes: selectivity between bacteria and eukaryotic cells. *J Antimicrob Chemother* 2016;71(6):1547-1555.
 23. Bazargani MM, Rohloff J. Antibiofilm activity of essential oils and plant extracts against *Staphylococcus aureus* and *Escherichia coli* biofilms. *Food Control* 2016;61:156-164.
 24. Leonard PG, Bezar IF, Sidote DJ, Stock AM. Identification of a hydrophobic cleft in the LytTR domain of AgrA as a locus for small molecule interactions that inhibit DNA binding. *Biochemistry* 2012;51(50):10035-10043.
 25. Trott O, Olson AJ. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J Computat Chem* 2010;31(2):455-461.
 26. Morris GM, Goodsell DS, Halliday RS, Huey R, Hart WE, Belew RK, et al. Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *J Comput Chem* 1998;19:1639-1662.
 27. Laskowski RA, Swindells MB. LigPlot+: multiple ligand-protein interaction diagrams for drug discovery. *J Chem Inf Model* 2011;51(10):2778-2786.
 28. Mhalla D, Bouaziz A, Ennouri K, Chawech R, Smaoui S, Jarraya R, et al. Antimicrobial activity and bioguided fractionation of *Rumex tingitanus* extracts for meat preservation. *Meat Sci* 2017;125:22-29.
 29. Akinpelu DA, Odewade JO, Aiyegoro OA, Ashafa AO, Akinpelu OF, Agunbiade MO. Biocidal effects of stem bark extract of *Chrysophyllum albidum* G. Don on vancomycin-resistant *Staphylococcus aureus*. *BMC Complement Altern Med* 2016;16:105.
 30. Desbois AP, Smith VJ. Antibacterial free fatty acids: activities, mechanisms of action and biotechnological potential. *Appl Microbiol Biotechnol* 2010;85(6):1629-1642.
 31. Negi PS. Plant extracts for the control of bacterial growth: efficacy, stability and safety issues for food application. *Int J Food Microbiol* 2012;156(1):7-17.
 32. El-kalyoubi S, Fatmah Agili F, Youssif S. Novel 2-thioxanthine and dipyrimidopyridine derivatives: synthesis and antimicrobial activity. *Molecules* 2015;20(10):19263-19276.
 33. Sanchez E, Rivas Morales C, Castillo S, Leos-Rivas C, Garcia-Becerra L, Ortiz Martinez DM. Antibacterial and antibiofilm activity of methanolic plant extracts against nosocomial microorganisms. *Evid Based Complement Alternat Med*. 2016;2016:1572697.
 34. Aqil F, Ahmad I, Owais M. Evaluation of anti-methicillin-resistant *Staphylococcus aureus* (MRSA) activity and synergy of some bioactive plant extracts. *Biotechnol J* 2006;1(10):1093-1102.
 35. Betoni JE, Mantovani RP, Barbosa LN, Di Stasi LC, Fernandes Junior A. Synergism between plant extract and antimicrobial drugs used on *Staphylococcus aureus* diseases. *Mem Inst Oswaldo Cruz* 2006;101(4):387-390.
 36. Klastersky J, Husson M. Bactericidal activity of the combinations of gentamicin with clindamycin or chloramphenicol against species of *Escherichia coli* and *Bacteroides fragilis*. *Antimicrob Agents Chemother* 1977;12(2):135-138.
 37. Jawetz E, Gunnison J, Speck R, Coleman V. Studies on antibiotic synergism and antagonism: the interference of chloramphenicol with the action of penicillin. *AMA Arch Intern Med* 1951;87(3):349-359.
 38. Nemeth J, Oesch G, Kuster SP. Bacteriostatic versus bactericidal antibiotics for patients with serious bacterial infections: systematic review and meta-analysis. *J Antimicrob Chemother* 2015;70(2):382-395.
 39. Soriano A, Marco F, Martinez JA, Pisos E, Almela M, Dimova VP, et al. Influence of vancomycin minimum inhibitory concentration on the treatment of methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Infect Dis* 2008;46(2):193-200.
 40. Tsuchiya H, Sato M, Miyazaki T, Fujiwara S, Tanigaki S, Ohyama M, et al. Comparative study on the antibacterial activity of phytochemical flavanones against methicillin-resistant *Staphylococcus aureus*. *J Ethnopharmacol* 1996;50(1):27-34.
 41. Cushnie TT, Lamb AJ. Recent advances in understanding the antibacterial properties of flavonoids. *Int J Antimicrob Agents* 2011;38(2):99-107.
 42. Ohta S, Shiomi Y, Kawashima A, Aozasa O, Nakao T, Nagate T, et al. Antibiotic effect of linolenic acid from *Chlorococcum* strain HS-101 and *Dunaliella primolecta* on methicillin-resistant *Staphylococcus aureus*. *J Appl Phycol* 1995;7(2):121-127.
 43. Huang CB, Alimova Y, Myers TM, Ebersole JL. Short- and medium-chain fatty acids exhibit antimicrobial activity for oral microorganisms. *Arch Oral Biol* 2011;56(7):650-654.
 44. Borges A, Abreu A, Malheiro J, Saavedra MJ, Simões M. Biofilm prevention and control by dietary phytochemicals. Microbial pathogens and strategies for combating them: science, technology and education. *Formatex Research Center, Badajoz*. 2013:32-41.